

LANIOTURDUS

Volume 54 No 1

March 2021



Journal of the Namibia Bird Club
<https://www.namibiabirdclub.org>

About the Namibia Bird Club

The Namibia Bird Club was founded in 1962 and has been active since then. The club's mission is to contribute to Namibian ornithology by, amongst other things, arranging regular birding outings, conducting bird ringing and atlasing excursions and educating the public about the value of birds. To achieve this, we organise monthly visits to interesting birding sites around Windhoek as well as regular visits to Avis Dam and the Gammams Water Care Works and occasional weekend trips further afield. Bird club members also participate in the African Waterbird Census twice a year.

Experienced birders are more than happy to help beginners and novices on these outings. If you have a transport problem or would like to share transport please contact a committee member. Depending on the availability of speakers and suitable material we present occasional lecture or video evenings at the Namibia Scientific Society premises. Members receive the bird club's journal, *Lanioturdus* and outings and events are advertised on the club's website www.namibiabirdclub.org.

The Namibia Bird Club is not affiliated to any global or regional organisation and relies entirely on members' subscriptions and donations to fund its activities.

The opinions expressed in this journal are those of the authors and not necessarily those of the Namibia Bird Club or its committee.



LANIOTURDUS

Vol. 54 (1) 2021

March 2021

<https://www.namibiabirdclub.org>

CONTENTS

KELLY M	Attracting insectivorous birds to the Namibian garden 1
DÜVEL S	Fork-tailed Drongo feeding on Scaly- feathered Weaver 5
CUNNINGHAM P	Southern Fiscal prey on tenebrionid beetles and Black-eared Sparrow-Lark in southern Namibia..... 6
KOLBERG H	Observations of African Fish Eagles, Saddle- billed Storks and Wattled Cranes during an aerial survey in north-eastern Namibia 10
BRYSON U	Birds in the hand: Ageing and sexing of Rosy- faced Lovebirds <i>Agapornis roseicollis</i> in Namibia..... 13
MEYER J	Bird names in the languages of Namibia 22
THOMSON N	Rarities and Interesting Sightings 27
KOLBERG H	Bird Atlas Update 36
KOLBERG H	Tracked Vultures 37

Cover photo Grey-backed Sparrow-Lark © Holger Kolberg

Attracting insectivorous birds to the Namibian garden

Mel Kelly

marisak@iway.na

All photographs are © Mel Kelly

Over time, a feeding station set up for wild birds in the garden can attract a great many different species (including striped grass mice). Often, individual birds become habituated to arriving at the exact time that seed dispensers and fruit supplies are replenished daily and will congregate a few moments beforehand, surrounding the site impatiently like over-anxious diners jostling for place at a popular café. On a summer's morning I've seen as many as 23 different bird species from my kitchen window on a property just outside Windhoek, the majority of them attracted by the various foodstuffs available at the feeding station in and under a *swarthaak* tree nearby.



Figure 1: Feeding post in use

In order to encourage the somewhat shyer insectivorous birds to visit, it's necessary to have a source of live

food of course and many birders have a mini 'mealworm farm' set up at home for this purpose, harvesting a handful of the grubs every day and letting the remainder pupate and then develop into the adult beetles that continue the breeding process (Gudrun Middendorff and Neil Thomson can give advice on setting one up at home and maybe even give you some of their excess stock.)

However, just as certain 'pushier' vegetarian species become familiar with the time(s) that meals are served and tend to dominate proceedings (Rosy-faced Lovebirds, I'm looking at you!), so individual birds will also tend to monopolise a live-food supply and – due to their more aggressive natures – prevent others from feeding. In fact, when mealworms are simply scattered on the ground, they will often be consumed not by their intended beneficiaries but by species such as the White-browed Sparrow-Weaver that are perfectly satisfied feeding from the seed dispenser ordinarily but which turn out to be pretty opportunistic and indiscriminate when it comes to quickly devouring anything else on offer.

One easy method for controlling how mealworms are dispensed – so that Marico Flycatchers, Familiar Chats, Short-toed Rock Thrushes, Crimson-breasted Shrikes and Cape Wagtails can take their turn – is to make a simple device that allows the mealworms to become available only sporadically, over a period of several hours. In this way, one or two may be eaten when the hordes descend to exhaust the seed, nut and fruit

supplies but the species that predominantly eat insects will soon learn to gravitate towards the supply over the entire course of the day, spotting whenever a mealworm becomes available and swooping down to grab it.

1. Select the place where you wish the insectivorous birds to feed – as they are fairly shy initially this should be maybe 15 - 20 metres from the house, preferably a level paved area or concrete slab so the liberated mealworms are easily seen and don't disappear into grass or gravel (mine is set up over a concrete bench). If you have pets or mongooses that enter the garden, I wouldn't recommend situating the 'feeding post' anywhere that will make the birds using it vulnerable to animals that might frighten them off (or worse).
2. Begin by placing a small handful of mealworms on the ground a few times a day at the chosen site so that the insectivorous birds in the area become familiar with the position of the planned feeding post, and come to realise that live food will be readily available throughout the daylight hours.
3. Once you have observed some insectivorous birds regularly visiting to feed on the mealworms you place on the ground, you can position your mealworm dispenser in the spot you have chosen. It may take only a couple of days for the birds in question to begin visiting since their eyesight is good and they spot their prey from quite a distance away, or it may take several weeks – especially if you don't have high trees or structures nearby that provide a vantage

point from which birds can observe the mealworms.



Figure 2: Mealworms in open pot

4. Buy a small (+/- 10cm diameter), semi-transparent, rigid plastic food-storage pot with a screw-fit lid – these are readily available in any supermarket for a few dollars (you can recycle old plastic containers that are used to sell dips etc. but the push-on lids tend not to be very secure and the thinner plastic soon cracks).
5. Heat a bradawl or small screwdriver over a naked flame and use it to melt 4 or 5 holes right through the base of your container, pushing from the inside outwards. Start off by making them *slightly* less than the estimated circumference of the average mealworm – if they struggle to crawl free you can always make the holes a little bit bigger.
6. Next, punch three holes in the lid using the same method to melt them through (if you just try to make holes by stabbing away at the plastic with an un-heated implement, it will split).



Figure 3: closed pot and hanging loop

7. Thread some ribbon, heavy-duty thread, or elastic through the holes in the lid to fashion a hanging loop, then attach this to a long piece of similar cord that will suspend the container over the spot you have chosen.
8. Push a long bamboo pole (+/-2 metres) deep into the ground or into the soil in a decent-sized pot at the site where you have been supplying the grubs. Then cut through an old wire coat-hanger and bend it open to make a straight 'arm' 30 – 50 cm long. Push one end 5 cm into the top of the bamboo pole then carefully bend the protruding length at right-angles over the place where you have been scattering the mealworms. This will form the perch for the birds to observe the supply of worms as they escape from the pot onto the ground. Once you are happy with the position of the arm, secure it to the top of the bamboo cane with some insulating tape or duct tape

so that it doesn't pivot around the pole.

9. Bend the free end of the wire into a closed loop to take the cord by which you will hang the container. Suspend the pot at the desired height, making sure that it doesn't catch on any nearby plants when it swings in the breeze, then cut off any remaining cord that's not needed once you have tied it firmly in place. You need to be able to reach up and unscrew the bottom portion of the pot daily so don't hang it too high if you are a shortish person (like me).



Figure 4: Suspended pot in situ

10. Finally, count out a few mealworms, place them inside the pot, screw the pot back onto the suspended lid, and check back a bit later to see how many have escaped. If none have emerged after an hour or so, you will need to increase the number of holes and/or make the holes bigger.

You are aiming to have maybe one grub fall out every $\frac{1}{4}$ hour eventually – although you may find that birds soon learn to come and grab any dangling from the holes before they even get a chance to tumble free. It's not an exact science as mealworm behaviour is a bit of an unknown quantity and in my experience those remaining in the pot tend to become a bit lethargic as the day wears on (regardless of the weather), so any that are still left in the container by early evening I then tip onto the ground so that they don't end up miserable in a confined space overnight.

11. Once you have the size and number of holes in the pot correctly calibrated, you can begin to add more substantial numbers of grubs, although be aware that in cooler weather their reproductive rate in the farm

slows down and you should not reduce the population to such a degree that you start to run out of stock.

You can, of course, modify the equipment for your particular circumstances and, for example, suspend the pot from a low branch in a suitable tree. I've also had success using a long, horizontal plastic tube that once held a poster, which gives a much bigger area of coverage of course, though you cannot see inside to monitor the supply of live-food remaining at any given time.

I'd be very interested to know if this technique works for you, and what species you are able to attract. Though Common Scimitarbills and Southern Pied Babblers are abundant where I live, neither species has visited the feeding post to date, though it hasn't been up for very long and they may yet begin to use it over time.

Fork-tailed Drongo feeding on Scaly-feathered Weaver

Sylvia Düvel

omambond@iway.na

All photographs are © Sylvia Düvel



At the beginning of August 2019 I heard a commotion in our garden. When I reached the tree with my camera, I saw a Fork-tailed Drongo

hacking away at the head of a Scaly-feathered Weaver.



After a time I checked again and both birds were gone. I assume that the drongo had eaten it all. According to Roberts', drongos do feed on small birds as well as insects, even fish and nectar.

Southern Fiscal prey on tenebrionid beetles and Black-eared Sparrow-Lark in southern Namibia

Peter Cunningham (Snr) and Peter Cunningham (Jnr)

pckkwrc@yahoo.co.uk

Southern Fiscal (*Lanius collaris*) have a catholic diet and although they are known predators of invertebrates there is often a paucity of data regarding specific species, especially when referring to beetles (Soobramoney *et al.* 2004, Hockey *et al.* 2005). References to beetles typically include generic terms such as “beetles”, “dung beetles”, “coleoptera”, “tenebrionid beetles” but are rarely species specific (Pearson 1962, Skead 1995, Kopij 1999, Soobramoney *et al.* 2004). However, references to birds are more comprehensive and include Quailfinch, Brubru, Cape Canary, Cape Sparrow, Grey Tit, Golden-breasted Bunting, Helmeted Guineafowl (chicks), House Sparrow, Levallant’s Cisticola, Mountain Wheatear, Speckled Pigeon (chicks), Stonechat and Wailing Cisticola, (Siegfried 1965, Ade 1978, Weaving 1980, Dean & Dean 1987, De Swart 1989, Claasen & Claasen 1989, Roos & Roos 1988, De Swart 1990, Skead 1995, Kopij 1999, De Swart 2016). Although passerine birds are only occasionally preyed upon by Southern Fiscal (Kopij 1999), these larger avifauna prey items may contribute significantly to their diet in terms of biomass (Soobramoney *et al.* 2004).

Prey items are often impaled or wedged on thorns, wood splinters, and barbs of wire fences (Hockey *et al.* 2005) which serves as a ‘storage’ function (Bevan & England 1969) or as a display for territorial advertisement and to attract females (Yosef & Pinchow 1989). However, Southern Fiscal do not cache as

much as their northern hemisphere counterparts (Harris & Arnot 1988).

Tenebrionid beetles

On 13 April 2020 we found a tenebrionid beetle – *Onymacris multistriata* (Haag-Rutenberg, 1875) – impaled on a *Boscia foetida* (smelly shepherd’s tree) shrub (at a height of 1.5m within a shrub with a total height of approximately 2m) by a Southern Fiscal on a farm approximately 70 km south of Grünau in the Karas Region, southern Namibia (28°16’12.7”S & 18°03’44.1”E; 740m) (Figure 1). *Onymacris multistriata* is viewed as a near endemic to Namibia (See: Namibia Biodiversity Database – www.biodiversity.org.na) and usually associated with sandy areas in southern Namibia (Irish *Pers. com.*) where it searches for food during the day (Louw & Seely 1982). As few insects are active during the activity periods of these beetles, they potentially have many predators (Duncan 2003).

On 6 September 2020 another tenebrionid beetle – *Somaticus aeneus* (Solier, 1843) – was found impaled on a different *Boscia foetida* shrub (at a height of 1.4m on the southern outer perimeter of the shrub with a total height of approximately 2.5m) (Figure 2). *Somaticus aeneus* (tar darkling beetle) are known to occur in south-eastern Namibia (See: Namibia Biodiversity Database – www.biodiversity.org.na), although they are more widespread in South Africa, where they scavenge on plant and animal material (Picker *et al.* 2019).

Black-eared Sparrow-Lark

On 21 April 2020 we found a Black-eared Sparrow-Lark – *Eremopterix australis* – impaled on another *Boscia foetida* shrub (at a height of 1m on the northern outer perimeter of the shrub with a total height of approximately 2.25m) on the same farm although approximately 4km from the beetle site (Figure 3). The head of the unfortunate bird was not present and probably consumed prior to impalement. Black-eared Sparrow-Larks are nomadic and have only been observed on the farm after above average rainfall events with breeding confirmed during April 2020 (In prep).



Figure 3: Black-eared Sparrow-Lark impaled on a twig on the outer perimeter of a *Boscia foetida*.



Figure 1: *Onymacris multistriata* impaled on *Boscia foetida* (smelly shepherd's tree) twig.



Figure 2: *Somaticus aeneus* impaled on a twig on the southern side of a *Boscia foetida*.

Although the actual impaling processes for all three species were not observed, this is inferred, as Southern Fiscal are resident on the farm. Previously they have been observed preying on unidentified invertebrates, an unidentified yellow scorpion (probably *Parabuthus brevimanus*) and even a western three-striped skink (*Trachylepis occidentalis*) (Cunningham 2018).

Southern Fiscal are often seen on the farm, although these were the first impaled invertebrate and bird prey items encountered. The last few years on the farm have been associated with below average annual rainfall with relatively few arthropods observed during this period. Under these circumstances the birds probably consume what is caught rather than cache their prey.

Although the breeding season for Southern Fiscal is typically between August and January (Tarboton 2001) and between August and June in Namibia (Jarvis *et al.* 1999), these “kill and display” of the beetles and Black-eared Sparrow-Lark could serve a social function. However, the placement of the dead *Onymacris multistriata* within the *Boscia foetida* shrub contradicts this, although it is not clear how Southern Fiscal view their surroundings.

Southern Fiscal prey on a wide variety of arthropods, mostly insects, with Coleoptera the third most important group after Hymenoptera and Orthoptera, while birds only account for between 1-2% of all prey items, as determined in KwaZulu Natal, South Africa (Soobramoney *et al.* 2004). Kopij (1999) indicates that tenebrionidae and scarabacidae form the largest (and equal) part of coleoptera prey in the Free State, South Africa. In southern Namibia, it is expected that they would prey on anything they are capable of catching, especially in a marginal environment and under drought conditions.

Nevertheless, this is a first record of *Onymacris multistriata*, *Somaticus aeneus* and Black-eared Sparrow-Lark as prey by Southern Fiscal, as far as we could determine.

Acknowledgements

John Irish is acknowledged and thanked for the identification of *Onymacris multistriata* from a photograph.

References:

Ade, B. 1978. Fiscal shrike killing house sparrow. *Honeyguide* **93**: 41. In: Hockey, P.A.R., Dean, W.R.J., Ryan, P.G. (eds.) 2005. *Roberts – Birds of Southern Africa*, VIIth ed. The Trustees of the John Voelcker Bird Book Fund, Cape Town.

Bevan, G. and England, M.D. 1969. The impaling of prey by shrikes. *British Birds* **62**: 192-199.

Claasen, J. and Claasen, R. 1989. Voedsel van fiskaallaksman. *Mirafra* **6**: 72.

Cunningham, P.L. 2018. *Trachylepis occidentalis* (Peters 1867). Western three-striped skink. Avian Predation. *African Herp News* **67**: 32-34.

Dean, S.J. and Dean, W.R.J. 1987. Fiscal shrike predation on mountain chat. *Promerops* **180**: 17-18.

De Swart, D. 1989. A wailing cisticola caught by a fiscal shrike. *Mirafra* **6**: 45.

De Swart, D. 1990. Bontrokkie val ten prooi van 'n laksman. *Mirafra* **7**: 49.

De Swart, D. 2016. Lesser grey shrike and common fiscal impaling prey. *Biodiversity Observations* **7.57**: 1-2.

Duncan, F.D. 2003. The role of the subelytral cavity in respiration in a tenebrionid beetle, *Onymacris multistriata* (Tenebrionidae: Adesmiini). *Journal of insect physiology* **49**: 339-346.

Harris, T. and Arnot, G. 1988. *Shrikes of southern Africa*. Struik Winchester, Cape Town. In: Hockey, P.A.R., Dean, W.R.J., Ryan, P.G. (eds.) 2005. *Roberts – Birds of Southern Africa*, VIIth ed. The Trustees of the John Voelcker Bird Book Fund, Cape Town.

Hockey, P.A.R., Dean, W.R.J. and Ryan, P.G. (eds.) 2005. *Roberts – Birds of Southern Africa*, VIIth ed. The Trustees of the John Voelcker Bird Book Fund, Cape Town.

Jarvis, A.M., Robertson, A., Brown, C.J. and Simmons, R.E. 1999. *Namibian avian database*. National Biodiversity Programme, Ministry of Environment and Tourism, Windhoek.

Kopij, G. 1999. Breeding ecology of the fiscal shrike *Lanius collaris* in a peri-urban environment in Bloemfontein, South Africa. Navorsinge van die Nasionale Museum Bloemfontein **15**: 45-63.

- Louw, G.N. and Seely, M.K. 1982. *Ecology of Desert Organisms*. Longman, London.
- Pearson, T.B. 1962. Foods and feeding (18). *Lammergeyer* **2**: 65. In: Hockey, P.A.R., Dean, W.R.J., Ryan, P.G. (eds.) 2005. *Roberts – Birds of Southern Africa*, VIIth ed. The Trustees of the John Voelcker Bird Book Fund, Cape Town.
- Picker, M., Griffiths, C. and Weaving, A. 2019. *Field guide to insects of South Africa*. Struik Nature, Cape Town.
- Roos, L. and Roos, M. 1988. Do fiscal shrikes feed on their stored food? *Mirafra* **5**: 28-29.
- Siegfried, W.R. 1965. Fiscal shrike attacking young guineafowl. *Ostrich* **36**: 224.
- Skead, C.J. 1995. Life-history notes on East Cape bird species, 1940-1990. **Vol. 1**, Algoa Regional Services Council, Port Elizabeth. In: Hockey, P.A.R., Dean, W.R.J., Ryan, P.G. (eds.) 2005. *Roberts – Birds of Southern Africa*, VIIth ed. The Trustees of the John Voelcker Bird Book Fund, Cape Town.
- Soobramoney, S., Downs, C.T. and Adams, N.J. 2004. Variability in foraging behaviour and prey of the common fiscal shrike, *Lanius collaris*, along an altitudinal gradient in South Africa. *Ostrich* **75(3)**: 133-140.
- Weaving, A. 1980. Predation by fiscal shrike. *Honeyguide* **101**: 23.
- Yosef, R. and Pinchow, B. 1989. Cache size in northern shrikes influences female mate choice and reproductive success. *Auk* **106**: 418-421.

Observations of African Fish Eagles, Saddle-billed Storks and Wattled Cranes during an aerial survey in north-eastern Namibia

Holger Kolberg
holgerk@afol.com.na

The Ministry of Environment, Forestry and Tourism conducted aerial surveys on the river systems in north-eastern Namibia in October and November 2020. The main aim of the survey was to count wetland dependent mammals and crocodiles but the opportunity was used to also count African Fish Eagles (*Haliaeetus vocifer*), Saddle-billed Storks (*Ephippiorhynchus senegalensis*) and Wattled Cranes (*Grus carunculata*).

African Fish Eagles are considered a vulnerable species in Namibia due to over-fishing, pesticide abuse and degradation of riverine habitat. A total of 36 fish eagles were seen on the Okavango River, of these 28 (77%) were in the Bwabwata NP. On the rivers in the Zambezi Region, 140 fish eagles were recorded, 74 (52%) of these in protected areas (Table 1). Figures 1 and 2 show the spatial distribution of fish eagle sightings.

Table 1: Numbers of Fish Eagles, Saddle-billed Storks and Wattled Cranes recorded on the various river systems. Numbers are presented as inside parks/outside parks.

	Okavango	Kwando	Nkasa Lupala	Linyanti/ Liambezi	Zambezi/ Chobe	Total Zambezi Rivers
African Fish Eagle	28/8	14/2	60/0	0/37	0/27	74/66
Saddle-billed Stork	0	4/1	12/0	0/2	0/4	16/7
Wattled Crane	27/0	16/0	9/0	0	0	25/0

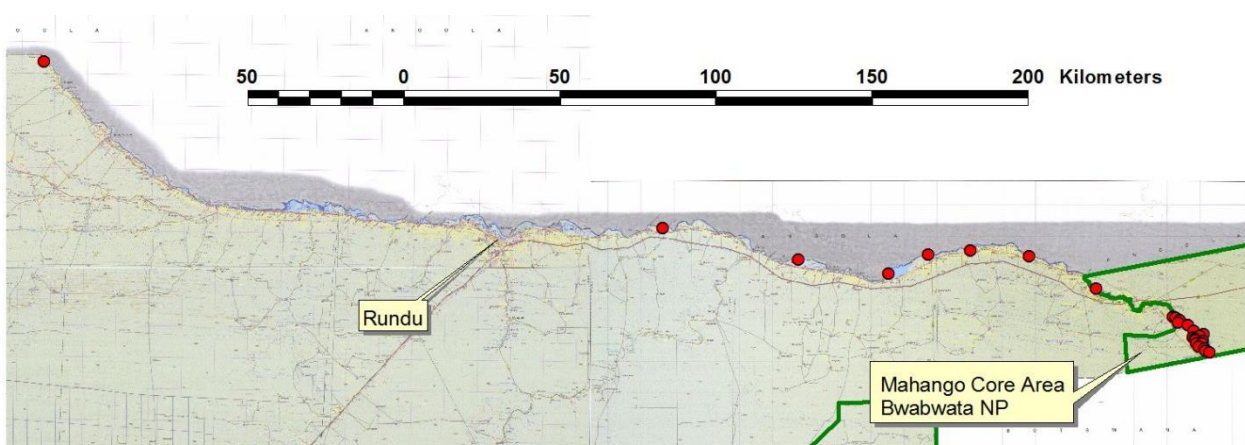


Figure 1: African Fish Eagle sightings (red dots) on the Okavango River (green lines = park boundaries).

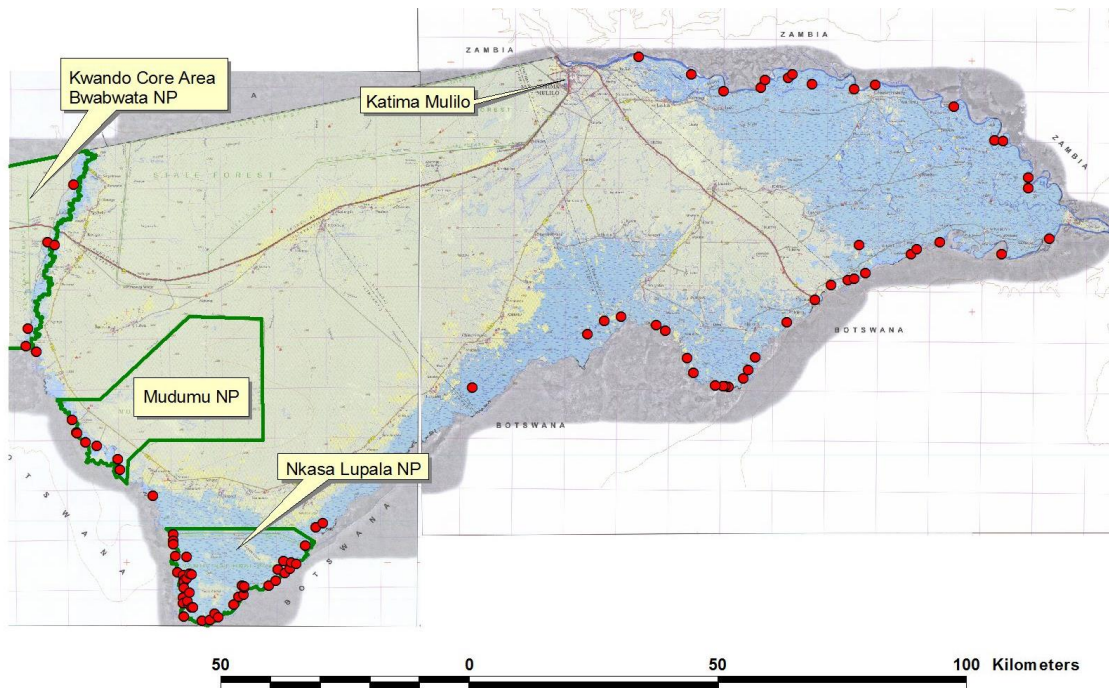


Figure 2: African Fish Eagle sightings (red dots) on the Zambezi Region rivers (green lines = park boundaries).

Saddle-billed Storks are regarded as endangered in Namibia due to increased human use of water and declining rainfall. No Saddle-billed Storks were seen on the Okavango but 23 in the Zambezi Region. Only

seven birds were recorded outside of protected areas, i.e. 16 (69%) of them were seen within the parks, the majority in Nkasa Lupala NP. Figure 3 shows the spatial distribution of Saddle-billed Stork sightings.

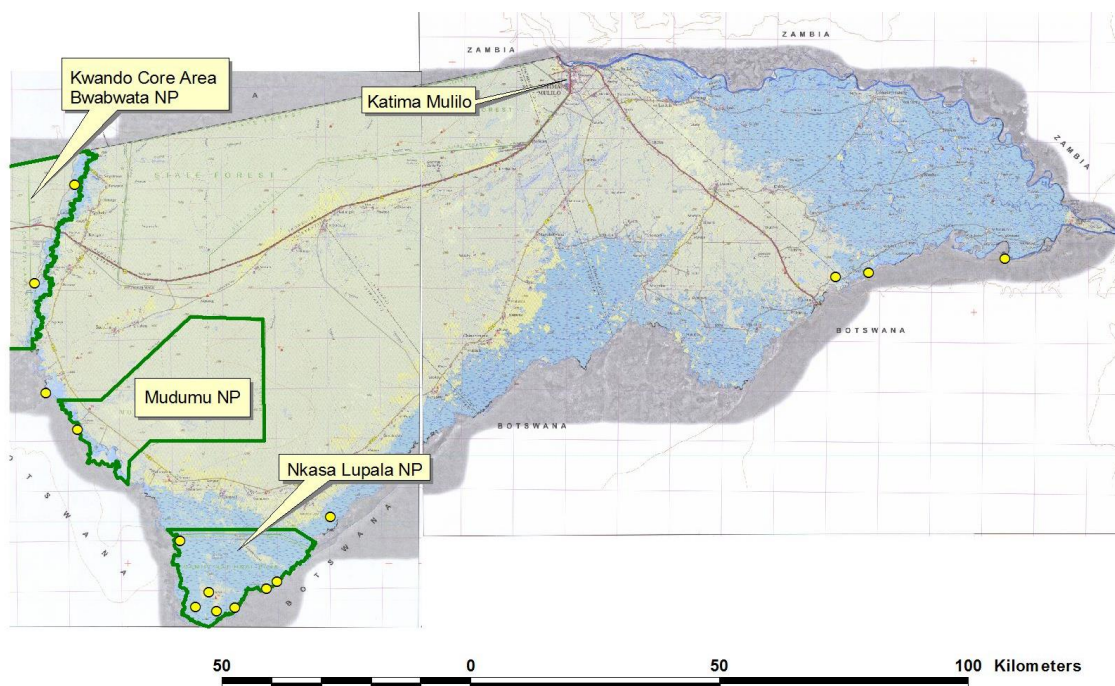


Figure 3: Saddle-billed Stork sightings (yellow dots) on the Zambezi Region rivers (green lines = park boundaries).

Wattled Cranes are also an endangered species in Namibia due to wetland degradation, grass burning and disturbance. On the Okavango River all 27 Wattled Cranes were found within the Bwabwata NP and

in the Zambezi Region too, all cranes were encountered in protected areas. Figures 4 and 5 show the spatial distribution of Wattled Crane sightings.

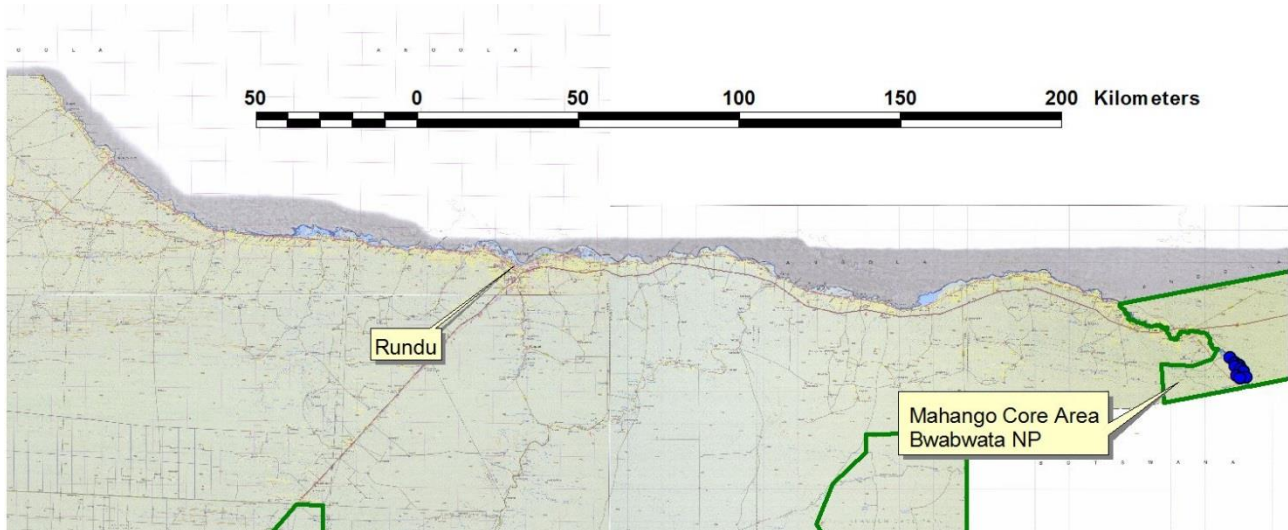


Figure 4: Wattled Crane sightings (blue dots) on the Okavango River (green lines = park boundaries).

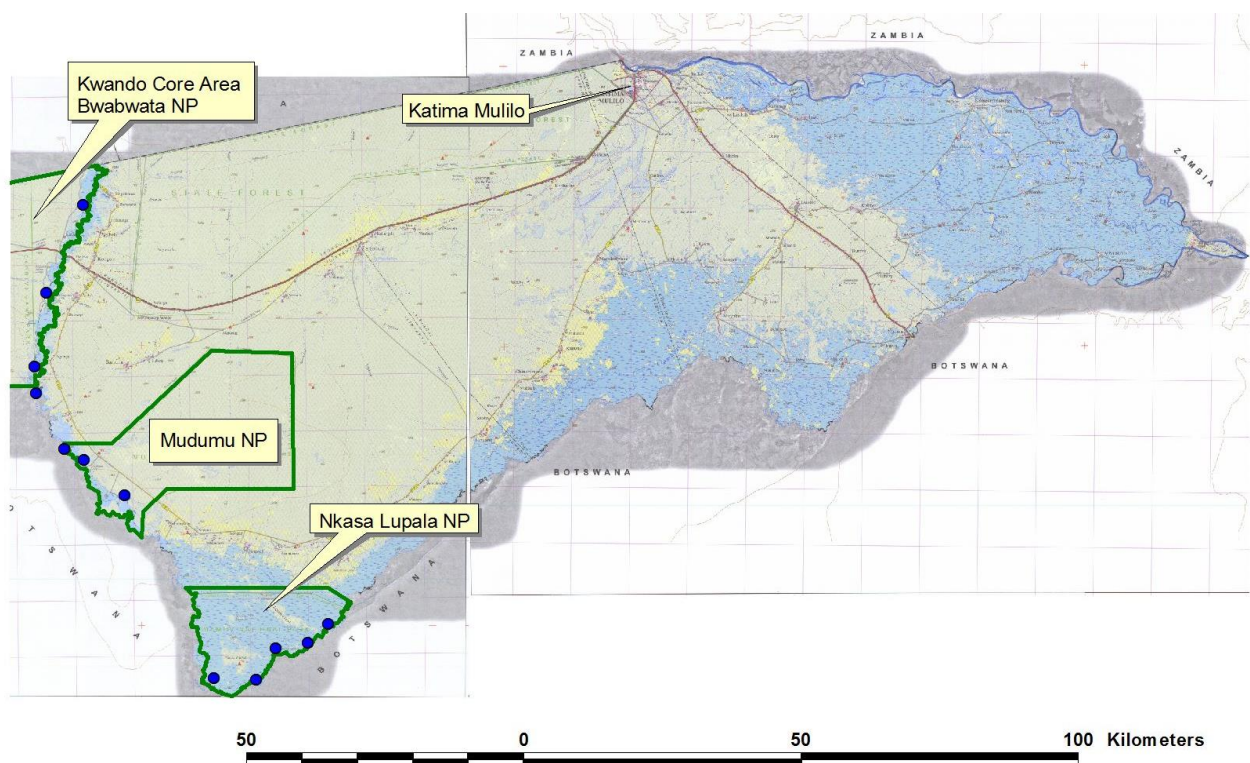


Figure 5: Wattled Crane sightings (blue dots) on the Zambezi Region rivers (green lines = park boundaries).

Birds in the hand: Ageing and sexing of Rosy-faced Lovebirds

***Agapornis roseicollis* in Namibia**

Ursula Bryson
ursula@thomas-bryson.de

In a discussion with another bird ringer the question arose, how to determine the gender and age of Rosy-faced Lovebirds. Breeders can rely on genetic testing to determine the sex of Rosy-faced Lovebirds, but bird ringers? We never had enough guidance and confidence to do so. The standard literature remains vague about the differences. We were astounded how very little information exists about the species in the wild. More concrete information can be found in breeding manuals or reports from the laboratory (Dilger 1960) which result from long term daily observations.



Figure 1: Rosy-faced Lovebirds at an artificial drinking well, Waterberg Plateau. Photo courtesy of Artur Bujanowicz.

Is a valid distinction possible by the colouration of the plumage? Are there other reliable criteria, when you watch a bird or have it in the hand just for a few minutes? Extensive study of the specific literature and interchange with breeders, researchers and geneticists confronted us with an unimagined complexity, and as always, we were left with more questions than before.

Sexing of Rosy-faced Lovebirds in their habitat

It is the behaviour that best tells the sexes apart: In the event of breeding, only the females build the nest and will sit on the eggs (the latter at least in captivity). It is they who carry sticks, grass, strips of bark and other nesting material to the nest. This is done by tucking it under the feathers of her rump and lower back (Forshaw 1989), while the male might prepare nest material but would not deliver it to the nest (Schwichtenberg 1973, p. 56). In captivity, same sex pairs have also been observed, especially in young birds or when a partner of the other sex is not available (Dilger 1960, p. 667). Does this occur also in the wild?



Figure 2: Presumably: male on the nest, female inside. Photo courtesy of Eckart Demasius.

And the plumage?

Described as pale grass green, slightly darker above with bright blue rump, peach pink face and upper chest, and darker reddish crown, it shows a substantial variation in its plumage colouration (see below). When two birds sit together, as pairs often do to preen each other, it is said that the slight sexual dimorphism can be observed: The pink of the head of the female is described as paler

and to less extensive (Schwichtenberg 1973, p. 15). On suitable pictures such plumage differences become visible. Alas, I learned by talking to breeders, curators, geneticists and ornithologists that these criteria give no definite certainty of the evidence of gender.



Figure 3: The bird on the left shows darker, broader cheeks vs. the one on the right with rosy-salmon face under the redder frons. Is this a difference in sexes, an age criterion or a result of breeding selection? In both individuals the extent of the red reaches down to the chest. Windhoek. Photo courtesy of Eckart Demasius.

Cheeks and eyes

You find a grey area of variable extent behind the pinkish cheeks in both sexes. Rosy-faced Lovebirds do not present white skin around the eye, as depicted in some of the literature, but just a thin ring of tiny white feathers around bare pale skin. They are thus unlike the group of "Agapornidae with white eye-ring", which includes the Yellow-collared *Agapornis personata*, Fischer's *A. fischeri*, Black-cheeked *A. nigrigenis* and Nyasa (Lilian's) Lovebird *A. lilianae*. These four species, esteemed by breeders, are wide spread as pets and escapees have established feral populations in some cities of South Africa, where they easily interbreed with escaped or released Rosy-faced Lovebirds.

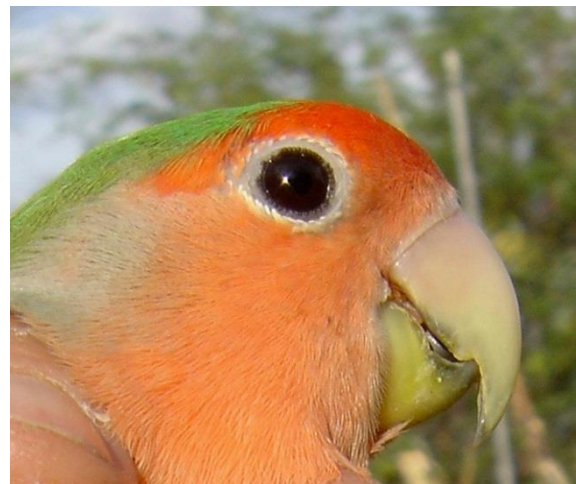


Figure 4: Face of a wild Rosy-faced Lovebird. The bare skin around the eye is flesh-coloured and encircled by small white feathers. From the edge of the Namib desert, Farm Sphinxblick, Namibia. Photo by UB.



Figure 5: Face of a wild Black-cheeked Lovebird. The broad circle around the eye is of white skin. Zambia. Photo courtesy of John Caddick.

Face and chest

The extent of the rosy colour in some individuals reaches down from the face to the chest and diffuses into the green while in others it terminates in an almost clear line slightly below the throat. Is this a consistent feature for sexing?

Again, no conclusive determination of sex is possible through this criterion.

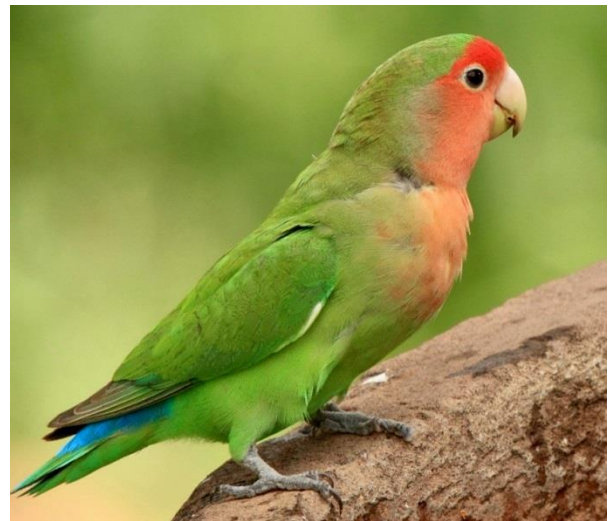


Figures 6 and 7: In some individuals the rosy mask is clearly confined under the throat. In others the peach/pink/rosy area diffuses into the chest. See also the difference between the shades of red in the face. (Photo top courtesy of Kevin Campher, Windhoek. Photo bottom by UB, Farm Sphinxblick.)

Shades of green

The body and the wings of some individuals seem to be almost uniformly coloured, while others display a clearly contrasting darker colouration of wing and back compared to the flanks and underparts. Looking through hundreds of pictures of wild birds, for plumage colour and moult features, beak colour, "tooth" shape and abrasion, quality of cere and the dorsal part of the bill, among others,

the question arises, whether the darkening might be a sign of age. This also includes the rump becoming a darker and deeper blue. Or are the contrasting colours of upper- and underparts an expression of the so-called "dark factor", a genetic disposition, which then shows more clearly in adult birds?



Figures 8 and 9: These two birds are from the same population at the Waterberg, their pictures were taken at the same occasion. They are wild and thus supposedly without human breeding interference. The difference between the extent of the rosy chest is striking, but also the colour difference between underparts/flanks and the wing (discussion see below). Photos courtesy of Tadeusz Rosinski.

A precise description of the nestling and juvenile plumage is missing. In the literature, it is called vaguely "as adult", with paler or duller colouration in the rose parts (Fry et

al. 1988; Perrin 2005), only Radtke (1984, p. 39) describes it as "similar but paler in all colours". More research is needed for the detailed documentation of the colouration of juvenile plumage, considering also the aspects of dark (and other) genetic factors, of escapees and of the influence of possible hybrids.

For example, comparing Figure 10 with Figure 11, the juvenile shows a light olive colouration. After a complete post-juvenile moult the plumage turns into a light green. We were wondering, whether in more mature birds the wings become even darker, or whether - again - the dark factor is showing.

See Figure 10 to 12 of three Rosy-faced Lovebirds with different shades of green, from the same photographic series taken by two co-photographers.

And then

We have, though, to take into consideration, that a genetic "dark factor", even in wild Namibian birds, has been observed which might show in a plumage of visibly darker or olive-green colour variation (van Den Abeele, pers. comm. 2020). And some individuals show brighter colours than those of their group like the individual in Figure 16. More research is needed to understand these variations in the wild.

Other genetically based colour variations have been found in the wild, like yellow or turquoise, though rarely. (See an overview of the recorded lovebirds of different species in van der Zwan et al. 2019).



Figure 10: Juvenile bird of a wild population, recognizable by the dark beak, the undefined pale face mask and the lack of red frons. Photo courtesy of Tadeusz Rosinski.



Figure 11 and 12: The plumage of the individual above shows almost the same brightness on the wing and the underparts, while the individual below has a distinctly darker wing and back compared to the flanks and underbelly, and a darker, more intensely coloured rump. Is this contrast of the latter a sign of age or the expression of a "dark factor"? Photos courtesy of Artur Bujanowicz. Waterberg Plateau.



Figure 13 and 14: Two individuals from the same population. The top bird in "normal" green, the bottom bird darker and with more olive tinge and yellowish flanks, Ugab Terrace Lodge. Photos courtesy of Olivier Laporte.

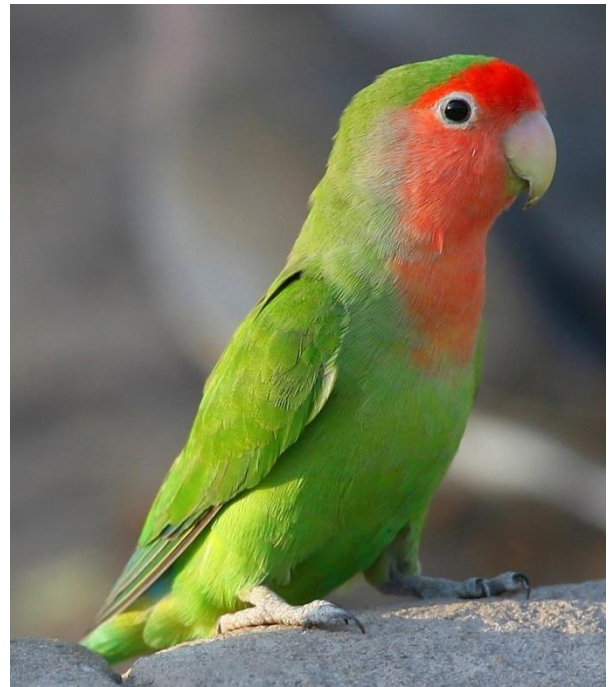


Figure 15: Rosy-faced Lovebird with intense red frons and strong rosy mask on bright green body, Roy's Camp. Photo courtesy of Bruce Robinson.

In the collection of the Ditsong National Museum of Natural History in Pretoria I found a "mutant", as it was labelled, from Namibia, of yellow plumage on the body and wing and salmon coloured head and face.



Figure 16: Colour "mutation" of a wild Rosy-faced Lovebird from Namibia, collected in April 1954. Ditsong National Museum of Natural History, Pretoria.

In February 2019, three different coloured juveniles of the same age (clutch?) were photographed in Henties Bay. The genetic lines are

determined by dominant, recessive and co-dominant genes, thus variations of siblings in the same brood are possible.



Figure 17: Juvenile Rosy-faced Lovebirds. The individual in the back is showing turquoise, not green plumage. The bird in the middle has a tendency to plain olive colouration, while in the bird in the front a difference between a bright green flank and a more olive wing is visible. Henties Bay. Photo courtesy of Mich Coker.

In the hand

The birds caught for ringing are mostly not recognizable as belonging to a distinct partner. Thus the possibility of comparison between two wild birds of a pair is rare.

In the literature we find that wing and tail are slightly longer in males, taken from a small sample of 7 males and 8 females, the beak length from the cere to the tip is equal in both sexes (Fry et al. 1988, Vol. 3, p.18), while females, at least observed in captivity, tend to be slightly larger than males although this difference is not apparent to the eye and is only reflected in average weights (Dilger 1960, p. 654).

Mass

For the field work, the mass alone as a sexing criterion seems questionable, as the birds in the wild are subjected to most variable conditions. Drought or rainfall with

subsequent changes in food supply, differing habitat such as sub-desert steppe, farmland with crops and waterholes or cities with constant food on offer will affect the physical fitness and mass.

While Dilger (1960, p. 651) records for captive birds 55,4g for males (18) and 56g for females (20) (presumably these data are given in Forshaw (1998): 55,4 (18 male), 56 (24 female) and then quoted in Hockey 2005), Fry et al. (1988) give for 29 unsexed, presumably wild, individuals 46 to 63g with a median of 54g. Out of 223 (except for a very small number) unsexed individuals, which we recorded over a period of more than 15 years in different areas of Namibia, the range was quite comparable to these data with 44 to 67,3g with a median of 52,4g.

The question of gender cannot be answered by the mass considering the small difference and the number of variables. And big males and small females may occur.

Beak

The beak of the female is said to have a wider base than in males. The head of the female is also more rounded than that of the male which is more massive and angular (Schwichtenberg 1973, p. 15). I could not find published measurements, nor photos depicting clearly these specific features. It might be of interest for ringers in the long run to include in the collected data those of the size of the head and the base of the beak.

Pelvic bones

Breeders recommend to check the gender by the pelvic bones with the bird in the hand (de Grahl 1990, p.140), if genetic testing is not possible. After sexual maturity the position and form of the pelvic bones are palpably different between the

sexes. In females, these bones are mostly stronger, rounded and a few millimeters apart. Those of the males appear weaker, more pointed and quite close to each other (Schwichtenberg 1973, p. 45). Sexual maturity has been observed in one single male at about 80 days (Dilger 1960, p. 635). This has been recorded in the literature as the first breeding age. "However, most birds begin to behave sexually at a considerably later date; usually after the post-juvenal moult is complete at about four months" (ibid.).

The technique of checking on the pelvic bones is being applied in other birds such as raptors or galliformes. As ringers we would need some basic guidelines for this method.

The distanced position of the pelvic bones in females leads to a sitting posture where legs are further apart than in males (de Grahl 1990, p.140). Due to a number of circumstances (landing, movement, position) this can be used as only one sexing criterion of many.

Colours

To make things even more interesting, the colouration of Rosy-faced Lovebirds may depend on different factors like the freshness or the bleaching and wear of the plumage and genetically based colour variations.

Rosy-faced Lovebirds are valued easy-to-breed pets with several broods possible per year in captivity, a fact that protects the wild populations from a wider persecution for trade.

For breeders "beautiful" birds are more attractive as they get higher prices on the market. "Beautiful" means in this context colourful or exquisite and rare colouration, like yellow, turquoise and blue and

others, similar to Australian Budgerigars *Melanopsittacus undulatus*. These colour forms occur in the wild, although very rarely.

In the recent decades, the number of Rosy-faced Lovebirds escaped from aviaries seem to have risen and numerous individuals have been regularly sighted out of their natural range. Feral populations have spread not only in cities in southern Africa, mainly in the area of Johannesburg and Pretoria (Symes 2014, p. 239; SABAP2), but can be found on all suitable continents, including in several European countries around the Mediterranean, and in other areas (Collar and Boesman 2020, see world map). The species thrives especially well in the USA, mainly in Arizona, with numbers estimated at over 2,000 (Audubon Field Guide 2020; Corman 2005). As obvious descendants from aviary birds, there is a huge colour variation in the now wild living birds. Other Agapornidae have started feral populations around the globe and interbreed easily with Rosy-faced Lovebirds.

It seems that the newly established and growing populations in Namibia belong mainly to natural populations which have benefitted from increasing water availability in the arid areas through farming, and from increasing man-made structures for nesting (C. Brown, pers. comm. 2020).

Acknowledgement

I want to thank the following photographers for generously entrusting me their pictures: Artur Bujanowicz, Kevin Campher, Mich Coker, Eckart Demasius, Olivier Laporte, Rubén Portas, Bruce Robinson, Tadeusz Rosinski. All other photos taken by the author. I am grateful to Chris Brown for sharing his knowledge, Dirk van Den Abeele

for his comments, and Janine Dunlop, from the Niven Library of the FitzPatrick Institute of African Ornithology, University of Cape Town, and Eva Karl, from the Library of the Zoologische Staatssammlung Munich, for their precious work. My thanks go also to the Ministry of Environment, Forestry and Tourism of Namibia for granting me the bird ringing permit.



Figure 18: Rosy-faced Lovebirds gathering during the dry season near surface water. About 60 km west of Epupa, Kunene. Photo courtesy of Rubén Portas.

References

- African Bird Club Image Data Base (Afbid)
<https://www.africanbirdclub.org/afbid/search/browse/species/729>.
- Audubon Society 2020.
<https://www.audubon.org/field-guide/bird/rosy-faced-lovebird>.
- Bryson U 2021. *Agapornis roseicollis* Rosy-faced Lovebird. In: Dueker et al. A review of research and conservation of the lovebirds of Africa: knowledge gaps and opportunities. Ostrich: Journal of African Ornithology, in prep.
- Collar NJ 1997. Rosy-faced Lovebird. In: del Hoyo J, Elliott A, Sargatal J (eds). Handbook of the Birds of the World, Vol. 4, Sandgrouse to Cuckoos, p. 410. Lynx Edicions, Barcelona.
- Collar N, Boesman PFD 2020. Rosy-faced Lovebird (*Agapornis roseicollis*), version 1.0. In: del Hoyo J, Elliott A, Sargatal J, Christie DA, de Juana E (eds). *Birds of the World*. Cornell Lab of Ornithology, Ithaca, NY, USA. https://doi.org/10.2173/bow.peflov.01_.
- Corman TE 2005. Peach-faced Lovebird (*Agapornis roseicollis*). In: Corman T, Wise-Gervais C (eds). Arizona Breeding Bird Atlas. University of New Mexico Press, Albuquerque.
- de Grahl W 1990. Papageien. Lebensweise, Arten, Zucht. 9th ed. Eugen Ulmer, Stuttgart. (1984 in English: The Parrot Family. New York: Arco Publishing.)
- Dilger WC 1960. The comparative ethology of the African Parrot genus *Agapornis*. Z. Tierpsychol. 17: 649-685.
- Forshaw JM 1989. Parrots of the World. 3rd revised edition. Blandford, London.
- Fry CH, Keith S, Urban EK 1988. The birds of Africa. Vol. 3: pp. 18-19. Academic Press, London.
- Hoesch W 1955. Die Vogelwelt Südwestafrikas. SWA Wissenschaftliche Gesellschaft. N.N.
- Hoesch W, Niethammer G 1940. Die Vogelwelt Deutsch-Südwestafrikas, namentlich des Damara- und Namalandes. Journal für Ornithologie 88 (Sonderheft).
- Juniper T, Parr M 1998. A Guide to the Parrots of the World. Pica Press, Sussex.
- McCarthy E. 2006. Handbook of avian hybrids of the world: pp. 124-125. Oxford University Press, Oxford.

Macdonald JD 1957. Contribution to the Ornithology of Western South Africa. Results of the British Museum (Natural History) Western South Africa Expedition, 1949 - 1950. British Museum (Natural History), London.

Moreau RE 1948. Aspects of evolution in the parrot genus *Agapornis*. Ibis 90: 206-239, 449-460.

Perrin MR 2005. Rosy-faced Lovebird *Agapornis roseicollis*. In: Hockey PAR, Dean WRJ, Ryan PG (eds). Roberts birds of southern Africa, 7th edition: pp. 227-228. The Trustees of the John Voelcker Book Fund, Cape Town.

Radtke GA 1984. Unzertrennliche (Agaporniden). Haltung, Zucht und Farbspielarten. Kosmos-Vivarium in Farbe, Franckh, Stuttgart.

SABAP 2. Rosy-faced Lovebird (*Agapornis roseicollis*). South African Bird Atlassing Project. <http://sabap2.birdmap.africa/species/330>.

Schwichtenberg H 1973. Die Unzertrennlichen. *Agaporniden*. Neue Brehm-Bücherei 400, 6th ed., Wittenberg Lutherstadt.

Simmons, RE 1997. Psittacidae: parrots, parakeet and lovebirds. In: Harrison JA, Allan DG, Underhill LG, Herremans M, Tree AJ, Parker V, Brown CJ (eds). The Atlas of southern African birds, pp. 534-535. BirdLife South Africa, Johannesburg.

Symes CT 2014. Founder populations and the current status of exotic parrots in South Africa. Ostrich. 85(3): 235 - 244.

van Den Abeele D 2016. Lovebirds compendium, genus *Agapornis*: species, breeding, genetics, mutations. About Pets, The Netherlands.

van der Zwan H 2019. *De novo* sequencing, assembly and annotation of the *Agapornis roseicollis* genome to identify variants for the development of genetic screening tests. orcid.org 0000-0001-5288-7609, <http://hdl.handle.net/10394/33154>.

van der Zwan H, Visser C, van der Sluis R 2019. Plumage colour variations in the *Agapornis* genus: a review, Ostrich, 90:1, 1-10, DOI: 10.2989/00306525.2018.1540446.

Bird names in the languages of Namibia

Johan Meyer
meyer.ij@gmail.com

Namibia is a culturally diverse country. Although English is the only official language, there are nine recognised national languages and three recognised regional languages. These statuses mean these languages are taught in school and used in national broadcasting. Two of these languages, Afrikaans and German, are linked to Namibia's colonial

history before independence. The other languages are indigenous to the area. In Table 1 there is a summary of the languages of Namibia, their names in the language itself and their status. As explained in Table 1, for this study, English, Afrikaans, German and Tswana were not included.

Table 1: The languages of Namibia

Languages	Language name in the language itself	Status
Afrikaans ¹	Afrikaans	National
English ¹	English	Official
German ¹	Deutsch	National
Herero	Otjiherero	National
Jul'hoan	Jul'hoansi	Regional
Khoekhoegowab	Khoekhoegowab	National
Kwangali	Rukwangali	National
Kwanyama ²	Oshikwanyama	National
Lozi	Silozi	National
Manyo	Rumanyo	Regional
Mbukushu	Thimbukushu	Regional
Ndonga ²	Oshindonga	National
Tswana ³	Setswana	National

1. Afrikaans, German and English do not form part of this study as they are Indo-European languages and already have species-specific names for all the birds of Namibia.

2. Ovambo is divided into the two standardised versions Ndonga and Kwanyama for the purposes of this study.

3. Tswana is not treated in this article as it is part of a project on bird names in the Sotho languages.

All birds have a scientific name that is used internationally to identify a bird up to species level. These names are unique for each species, but in reality, these names also change over time. These names are used alongside bird names in various languages. Depending on the language, a certain level of standardisation has taken place. In English, the International

Ornithological Committee's (IOC) names are mostly used. Similar standardisation of bird names has occurred for languages like French, Spanish and German to name a few. This has been done for all the birds of the world. For some languages, like Afrikaans, a regional list of standardised names is used.

These names are species specific. This means that each species has its own unique name in that specific language. Historically, most languages only have generic names used for similar birds. These birds may not even be related, but since they are similar, they have the same name. Due to this, names have been artificially standardised in many languages. The first attempt for this for any African language was done by H. Kolberg for Herero. He made a list of species-specific names for some birds of Namibia in 1986. This list was not complete for all the birds of Namibia but showed that bird names could be standardised in an African language. Before this list, only general bird names were known and used.

The aim of this study was to list the names that were species specific for the birds of Namibia found in literature sources.

Method

This study consisted of an extensive literature search. Mainly dictionaries, but also other sources, were consulted. All bird names found were recorded to create a database. This database had a specific structure.

With each entry the following was recorded:

1. The name in the specific language
2. The name in the language found in source (mainly English, Afrikaans or German)
3. The source name
4. The family of birds to which the name belongs
5. The scientific name if it is found in source.

The names that were species specific, meaning they refer to just one species, were noted and put in a

separate database. This species-specific names database was analysed for this study.

A full list of the sources of the literature search is found at the end of the article.

Results and Discussion

A list of all the species-specific names can be found in Table 2. A summary of the analysis of this list can be found in Table 3. This includes the number of unique names as well as the number of species that had a name. Some species had more than one name. The percentage of Namibian birds that have a species-specific name is also indicated.

Table 2: Species-specific names found for the birds of Namibia (See pages 41 - 57 at the end of this publication)

Herero had the most names. This is due to the work of H. Kolberg in 1986. He produced a list of various species for which he standardised the names. Kwanyama had the least names, only 7 representing 7 species. This does not necessarily reflect the number of names existing in the language, but rather an indication of the amount and quality of the sources used. Only one dictionary was consulted in the study.

Herero also had the highest percentage of Namibian birds with species specific names, and Kwanyama had the lowest. The average percentage of birds with species-specific names was only 8.44%. Even Herero with the highest number of species-specific names only had 17.77%. On the other end of the spectrum Kwanyama had a mere 1.03%.

Table 3: A summary of the analysis of the species-specific names

Language	Total number of species-specific names	Number of species with species-specific names	Percentage of Namibian birds with species-specific names
Herero	143	121	17.77
Jul'hoan	70	58	8.52
Khoekhoegowab	75	48	7.05
Kwangali	58	55	8.08
Kwanyama	7	7	1.03
Lozi	104	81	11.89
Ndonga	52	42	6.17
Manyo	75	66	9.69
Mbukushu	43	39	5.73
Average	69.67	57.44	8.44

Unlike English, Afrikaans and German, the other Namibian languages are lacking in names for birds. It has to be taken into account that only species-specific names were taken into account. All the languages mentioned here still have many generic names, where one name is used in general for more than one species. Thus, there is a need to standardise the names using these generic names by adding descriptive parts.

Conclusion.

It can clearly be seen that much research is needed in regards to bird names in the national and regional languages of Namibia. More dictionaries need to be consulted. Field work also needs to be done in order to document bird names before they get lost, as younger generations normally do not know the names of birds in their language. The rich birdlife of Namibia can only be fully appreciated by all Namibians if they have names for all of the birds in their own language. This will also create a better awareness of the rich birdlife that Namibia has and will aid in the conservation of birds.

Bibliography

- Cwi, T.F., Jones, K. 2014. *Jul'hoan Children's Picture Dictionary*. University of KwaZulu-Natal Press, Pietermaritzburg, South Africa.
- Dickens, P. 2009. *English - Jul'hoan, English - Jul'hoan Dictionary*. Rüdiger Köppe Verlag, Cologne, Germany.
- Gestwicki, R. 1966. *An English - Herero, Herero - English Dictionary*. Anglican Church, Windhoek, Namibia.
- Haacke, W.G.H., Eiseb, E. 2012. *A Khoekhoegowab Dictionary with an English-Khoekhoegowab Index*. Macmillan Education Namibia, Windhoek, Namibia.
- Haacke, W.G.H., Eiseb, E., Gericke, C. 2010. *Khoekhoegowab-Afrikaans Glossarium*. Macmillan Education Namibia, Windhoek, Namibia.
- Jalla, A. 1936. *Dictionary of the Lozi Language*. The United Society for Christian Literature, London, United Kingdom.
- Kloppers, J.K., Nakare, D., Isala, L.M., Bredell, A.W. 1994. *Rukwangali - English, English - Rukwangali*

- Dictionary*. Gamsberg Macmillan, Windhoek, Namibia.
- Kolberg, H. 1986. Die Liste der Vogelnamen in Otjiherero. *Lanioturdus* Vol. 21 Nr. 11. Windhoek, Namibia.
- Legère, K., Munganda, R. 2004. *English - Thimbukushu Subject Glossaries*. Gamsberg Macmillan, Windhoek, Namibia.
- Maclean, G.L. 1993. *Roberts Birds of Southern Africa, Sixth Edition*. John Voelcker Bird Book Fund, Cape Town, South Africa.
- Möhlig, W.J.G., Shikaya-Mberema, K.P. 2005. *A Dictionary of the Rumanyo language*. Rüdiger Köppe Verlag, Cologne, Germany.
- Olpp, J. 1888. *Nama- Deutsches Wörterbuch*. Elberfeld, Germany.
- Rust, F.R. 1960. *Deutsch-Nama Wörterbuch*. Rheinischen Mission in Südwestafrika, Windhoek, Namibia.
- Seidel, A. 1891. *Praktische des Nama, des Otjiherero und des Oshindnoga*. A Hartleben's Verlag, Wien, Austria.
- Snyman, J.W. 1975. *Žul'hōasi Fonologie en Woordeboek*. A.A. Balkema, Cape Town, South Africa.
- Tobias, G.W.R., Turvey, B.H.C. 1971. *English-Kwanyama Dictionary*. Witwatersrand Univeristy Press, Johannesburg, South Africa.
- Viljoen, J.J., Amakali, P. 1984. *Oshindonga Woordeboek - Dictionary - Embwiitya*. Gamsberg, Windhoek, Namibia.
- Viljoen, J.J., Amakali, P., Namuandi, M. 2000. *Oshindonga/English Embwiitya - Dictionary*. Gamsberg Macmillan, Windhoek, Namibia.
- Viljoen, J.J., Kamupingene, T.K. 2012. *Otjiherero woordeboek - dictionary - embo romambo*. MacMillan Education Namibia, Windhoek, Namibia.
- Wandres, C. 1918. Alte Wortlisten der Hottentottensprache. *Zeitschrift für Kolonialsprache*, p. 26-42. Hamburg, Germany.
- Wandres, C. 1927. Tiernamen in der Nama- und Bergdama-Sprache. *Festschrift Meinhof*, p. 125-133. Hamburg, Germany.

Rarities and Interesting Sightings

Compiled by Neil Thomson
batqs@afol.com.na

A Karoo Thrush was photographed in the vicinity of the Bosua Pass on 13 November 2020 (Birds of Namibia Facebook post). This sighting is commensurate with the apparent northward range expansion of this species.

On 14 November 2020 Dayne Braine reported both an Elegant Tern and a Common Redshank at Mile 4 Saltworks.

Ulrich Hofmann saw a Western Yellow Wagtail at Farm Kakuse some 70 km north west of Tsumeb on 15 November 2020. This was the first time he had seen this species on the farm.

Swakopmund's Swamp Boubou was seen again on 16 November 2020 (Birding Namibia WhatsApp Group post) (see *Lanioturdus* 53 [4]).

On 21 November 2020 Ursula Bryson mist netted a Jameson's Firefinch on a farm near Witvlei. This bird was a very long way from its known range. Does this signal the start of another firefinch range extension? Or is this perhaps an escapee from an aviary somewhere in the district?



Figure 1: Jameson's Firefinch © Ursula Bryson

On the same day Dayne Braine saw two Peregrine Falcons plus a few Cape (Glossy) Starlings at Henties Bay.

Yellow-throated Leafloves were seen at Caprivi Houseboat Safari Lodge on 24 and 25 November 2020.

Mark Boorman reported an Elegant Tern at Mile 4 Saltworks on 24 November 2020. The bird was still present the following day when, in addition, two Common Redshanks were seen as well.

Calle Schlettwein observed a European Honey Buzzard in his Eros garden on 27 November 2020.

A Gouldian Finch (an Australian species and obviously an escapee) was seen in Pioneers Park on 27 November 2020 (Birds of Namibia Facebook post).

Piet van Rooyen reported a solitary Black Stork at a dam on a farm west of the Gamsberg on 27 November 2020.

Gudrun Middendorff and I found a pair of Greater Painted-snipe at a dam (totally devoid of waterside vegetation) at Farm Teufelsbach between Windhoek and Okahandja on 28 November 2020.

Another European Honey Buzzard was reported in Klein Windhoek by Meryll Butcher on 30 November 2020.

Michael Houlden reported a probable Hooded Vulture near Namutoni on 29 November 2020 and a single White-headed Vulture near Halali the following day.



Figure 2: Greater Painted-snipe © Neil Thomson



Figure 3: White-headed Vulture © Michael Houlden

Ursula Bryson found a Ruddy Turnstone at a farm dam near the Waterberg on 30 November 2020. This is a most unusual record as this species is rarely found inland.

Uschi Bauer reported another White-headed Vulture sighting from the Etosha National Park on 30 November 2020, this one from Salvadoria.

Uschi then found a Caspian Plover near Ombika on 02 December 2020.

Hugo Haussmann photographed a Village Indigobird in Otjiwarongo on 02 December 2020. Its host, the Red-billed Firefinch, has previously been reported from the area.



Figure 4: Village Indigobird © Hugo Haussmann

Franz Klein saw a Red-necked Phalarope and two Bank Cormorants at Mile 4 Saltworks on 23 December 2020.

A Western Osprey was seen by Dayne Braine flying over the Strand Hotel in Swakopmund on 23 December 2020.

Dayne Braine reported that Swakpmund's Swamp Boubou had moved to Rossmund Golf Course on 24 December 2020 and shortly thereafter it was seen again in Swakopmund raising the possibility that there may be two of these birds present at the coast.

On 26 December 2020 Eckart Demasius reported a Western Yellow Wagtail at Rietfontein and at least twenty Lesser Kestrels at Charitsaub in the Etosha National Park. The following day he found seven Lesser Kestrels at Charitsaub and on 28 December found a Black-winged Pratincole at Koinachas. On 29 December Eckart found five Black-winged Pratincoles at Andoni and saw a White-headed Vulture over the Andoni Plains. On 30 December he saw two White-headed Vultures at Rietfontein.



Figure 5: Western Yellow Wagtail © Eckart Demasius

Small numbers of Red-necked Phalaropes were reported at Mile 4 Saltworks and at Walvis Bay by Mark Boorman and Franz Klein between 23 December 2020 and 02 January 2021. In addition, they saw two Red Knots at Walvis Bay on 24 December and Terek Sandpipers on 24 and 28 December 2020 and 02 January 2021.

Madina Fourie saw a Black Cuckoo at Swartfontein Lodge on 27 January 2020. This seems fairly far south for this species.

Danie Brand found a Eurasian Hobby near Outjo on 30 December 2020 – again quite far south for the species.

Ursula Bryson saw seven Lesser Kestrels south of the Waterberg on 31 December 2020.

Curt-Ingo Sagell mentioned a number of sought-after birds seen in the vicinity of Katima Mulilo on 01 January 2021. These included Yellow-throated Leaflove, African Emerald Cuckoo, Broad-tailed Paradise Whydah, White-winged Widowbird and African Hobby.

There were several sightings of Levillant's Cuckoo in central Namibia around the end of the year with two reported at Nuabamis by

Len van Zyl on 25 December and another reported at Okahandja on 01 January 2021. In addition, we saw one at Farm Lichtenstein Süd south west of Windhoek on 02 January.

A Red-chested Cuckoo, a very uncommon bird in Namibia, was heard calling in Eros on 03 and 04 January but to the best of my knowledge no-one actually saw the bird.

Kobus Bekker reported an influx of male Chestnut Weavers in the Naukluft area on 05 January 2021. This location is far south for the species.

A Lesser Moorhen was seen at N/a'an ku sê east of Windhoek on 09 January 2021 and another was reported at Avis Dam on 17 January (Birding Namibia WhatsApp Group posts). These are very far south for this species.

A Senegal Coucal was reported on a farm about 70 km south east of Grootfontein on 11 January 2021. While there have been a number of sightings of this species in the Grootfontein/Tsumeb/Otavi/Kombat area, this one is considerably further south than most (Birding Namibia WhatsApp Group post).

Merryl Butcher reported another European Honey Buzzard in Klein Windhoek on 12 January 2021.

Nic Buys found a Dwarf Bittern at Avis Dam on 12 January 2021.

On the same day we found two female Greater Painted-Snipe on the northern part of Farm Teufelsbach – the first time this species has been recorded on this pentad in SABAP2.

A Woodland Kingfisher was seen and heard around the farm house at

Farm Teufelsbach between mid-January and early February 2021. Our camera trap at a nest box approximately 2.5 km away recorded one on 23 February 2021. Interestingly 2021 was the first year we failed to record Woodland Kingfisher at Farm Monte Christo since the species was first detected there in 2009.



Figure 6: Woodland Kingfisher – camera trap

On 20 January 2021 Wessel Swanepoel reported a single Karoo Thrush in the vicinity of the Safari Motel in Windhoek. The bird had been present since 16 January. (See also above).

At least two Woodland Kingfishers were found by Nic Buys at Midgard Resort east of Okahandja on 20 January 2021.

Dayne Braine saw a single Terek Sandpiper at Lover's Hill, Walvis Bay, on the same day.

More aliens on the loose came to light with the report on 21 January 2021 of a pair of Java Sparrows in the

vicinity of Café Anton in Swakopmund (Birding Namibia WhatsApp Group post).

On 22 January Christiane Maluche saw three Black-winged Pratincoles between Kalahari Anib Lodge and Mariental. This is very far south for this species.



Figure 7: Black-winged Pratincole © Christiane Maluche

Photographs from Omaruru of both Red-billed Firefinches and Village Indigobirds were posted on the Birds of Namibia Facebook page on 22 January 2021 confirming the range extension of these species into that area.

Also on 22 January 2021 Bernette Louw spotted a juvenile African Harrier-Hawk in Henties Bay. There are no SABAP2 records for this species at the coast.



Figure 8: Juvenile African Harrier-Hawk © Bernette Louw

Dirk Bockmühl saw a Yellow-crowned Bishop at Farm Krumhuk south of Windhoek on 23 January 2021 and Elsbeth Elblein reported one in Pioneer Park Extension 1 about the same time.

On the same day Ulrich Hofmann found a Slaty Egret and a White-faced Whistling Duck at Farm Kakuse north west of Tsumeb.



Figure 9: Slaty Egret © Ulrich Hofmann



Figure 10: White-faced Whistling Duck © Ulrich Hofmann

The highlight of the Namibia Bird Club outing to Farm Haris south west of Windhoek on 24 January 2021 was the sighting of a juvenile Saddle-billed Stork (the identification of which prompted quite some debate). Also of interest was a partially leucistic South African Shelduck seen there.



Figure 11: Saddle-billed Stork © Neil Thomson



Figure 12: Leucistic South African Shelduck © Neil Thomson

Wilfried Nauhaus observed another Lesser Moorhen on a farm some 35 km south of Gobabis on 25 January 2021 (see above).

Mark Boorman found a Red-necked Phalarope at Mile 4 Saltworks on 25 January 2021 and a Common Redshank in near full breeding plumage at the same location on 26 January.



Figure 13: Red-necked Phalarope © Mark Boorman



Figure 14: Common Redshank © Mark Boorman

Two Blue-billed (Hottentot) Teal were seen by Eckart Demasius at Mile 4 Saltworks on 26 January 2021. There do not seem to be any previous records for this species at this location.



Figure 15: Blue-billed Teal © Eckart Demasius

A Woodland Kingfisher was reported at Otjiwa south of Otjiwarongo on 01 February 2021 (Birding Namib WhatsApp group).

A very unusual sighting reported on the Birds of Namibia Facebook page was that of a Sabine's Gull feeding on a seal carcass near Pelican Point on 02 February 2021. This marine species is seldom seen from the shore (see also below).

Holger Kolberg found two Pink-backed Pelicans at Hardap Dam on 02 February 2021 and another two at Naute Dam two days later.



Figure 16: Pink-backed Pelican (left) and Great White Pelican © Holger Kolberg

Ursula Bryson advised of the presence of a White-backed Duck and breeding plumaged Yellow-crowned Bishops near Witvlei on 03 February 2021. The previous week Ursula had seen two Hadedda Ibises in the same area.

Gudrun Middendorff saw a European Honey Buzzard over Klein Windhoek on 04 February 2021.

The leucistic Red-knobbed Coot (see *Lanioturdus* 53 [3]) was seen again at Avis Dam in the course of the wetland bird count conducted there on 06 February 2021.

Dayne Braine found two Red-necked Phalaropes at Mile 4 Saltworks on 06 February 2021.

Ten Yellow-billed Storks were seen in the course of the wetland bird count conducted at Farm Monte Christo north of Windhoek on 07 February 2021.

The saga of wandering juvenile Saddle-billed Storks (see above) continued at Otjivero Dam on 08 February 2021 when two were seen in the course of the wetland bird count conducted there. In addition, no fewer than nine Yellow-billed Storks were also seen at this location.

Rather surprisingly Holger Kolberg found another Sabine's Gull ashore at Shearwater Bay, Lüderitz, on 09 February 2021 (see also above).



Figure 17: Sabine's Gull © Holger Kolberg

A Red-chested Cuckoo was seen in Love Street, Windhoek on 09 February 2021.

An Egyptian Vulture was seen near Duineveld waterhole in the west of the Etosha National Park on 11 February 2021 (Birding Namibia WhatsApp group post).

The wetland bird count at the Cape Cross Lagoons on 12 February 2021 turned up no fewer than five Red-necked Phalaropes while a further two were seen along with a Common Redshank at Walvis Bay on 13 February.

A Diederik Cuckoo was heard calling in Swakopmund on 13 February 2021.

Mark Boorman photographed a Greater Sand Plover at Sandwich Harbour on the weekend of 13-14 February 2021.



Figure 18: Greater Sand Plover © - Mark Boorman

Hugo Haussmann found an Amur Falcon some 60 km south of Karibib on 14 February 2021. This is rather far south for this species.



Figure 19: Amur Falcon © Hugo Haussmann

Dayne Braine reported a Franklin's Gull at Walvis Bay on 15 February 2021.

After a prolonged disappearance Swakopmund's Swamp Boubou was seen again on 18 February 2021, this time in the industrial area on the eastern side of the town.

While monitoring our nest boxes at Farm Teufelsbach on 18 February 2021 we found, inter alia, a Grey-headed Kingfisher, a Dwarf Bittern, a Greater Painted-Snipe and a juvenile Yellow-billed Stork.

A Yellow-billed Oxpecker was photographed by Jaco Muller at Farm Reese near Outjo on 18 February 2021.

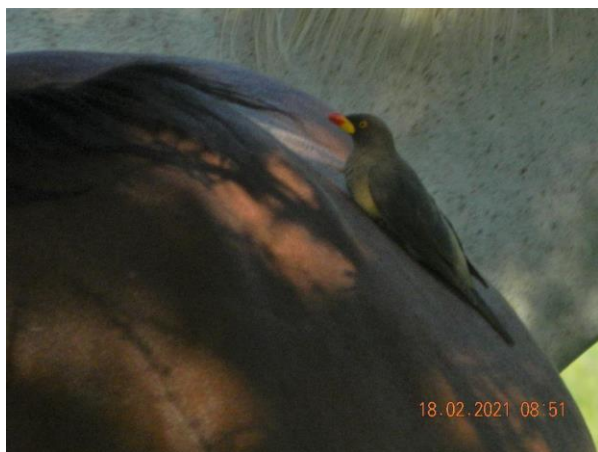


Figure 20: Yellow-billed Oxpecker © Jaco Muller

Hugo Haussmann reported a Western Osprey some 50 km south of Otjiwarongo on 19 February 2021

Madina Fourie reported another European Honey Buzzard in Olympia, Windhoek, on 20 February 2021.

Ulrich Hofmann saw a Malachite Kingfisher at Farm Kakuse on 21 February 2021. This bird is a long way from any SABAP2 record of this species.

In addition, Trevor Hardaker's SA Rare Bird News Reports mention the following rarities etc. seen in Namibia since early November 2020 :-

The long staying Ross's Turaco was seen again at Taranga Safari Lodge west of Rundu on 09 November 2020.

A Bradfield's Hornbill was seen at the plots about 8 km inland from Swakopmund on 14 November 2020 and again in the vicinity on 12 December. It was observed again at Rossmund golf course on 13 January 2021. (*This is in all likelihood the same bird reported in the general area before – see Lanioturdus 52[4] and 53[2] – NT*).

A European Honey Buzzard was reported in Cimbebasia, Windhoek, on 05 December 2020.

17 December 2020 produced a GIGA record when a single Northern Carmine Bee-eater was found amongst Southern Carmine Bee-eaters in the Buffalo Core Area of the Bwabwata National Park. This is a new species for the southern African subregion.

At least one Common Redshank was present at Mile 4 Saltworks on 18 December 2020.

Another European Honey Buzzard was seen east of Divundu on 19 December 2020 while one was reported in Windhoek on 23 December.

Swakopmund's Black Sparrow-Hawk was seen again on 20 December 2020.

Yellow-throated Leafloves were reported at Caprivi Houseboat Safari Lodge on a number of occasions between 23 December 2020 and 10 February 2021.

Three European Honey Buzzards were reported over Kleine Kuppe on 10 January 2021 and another was seen over Avis Dam on 13 January.

An African Skimmer was seen at Twee Palms in the Etosha National Park on 04 February 2021.

Another European Honey Buzzard was seen over Avis Dam on 06 February 2021.

SABAP2

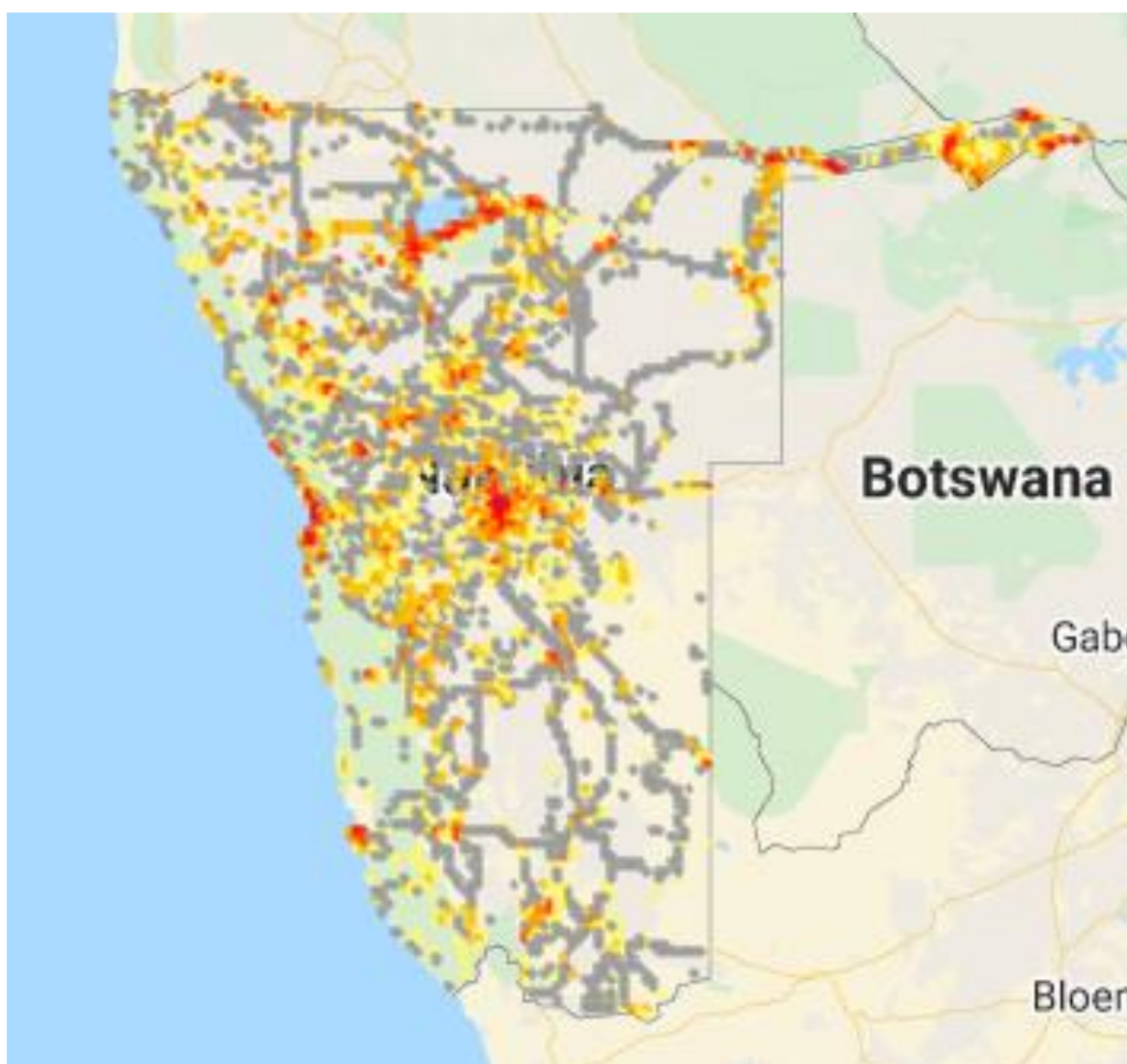


Bird Atlas Update

Holger Kolberg
holgerk@afol.com.na

1 809 pentads covered
640 species recorded
10 507 full protocol cards submitted

17.112% covered
445 species recorded in 2021
165 full protocol cards in 2021



Coverage map of Namibia; Grey squares represent *ad hoc* records, coloured squares full protocol cards – the colour indicates the number of times a pentad has been atlased.



Tracked Vultures

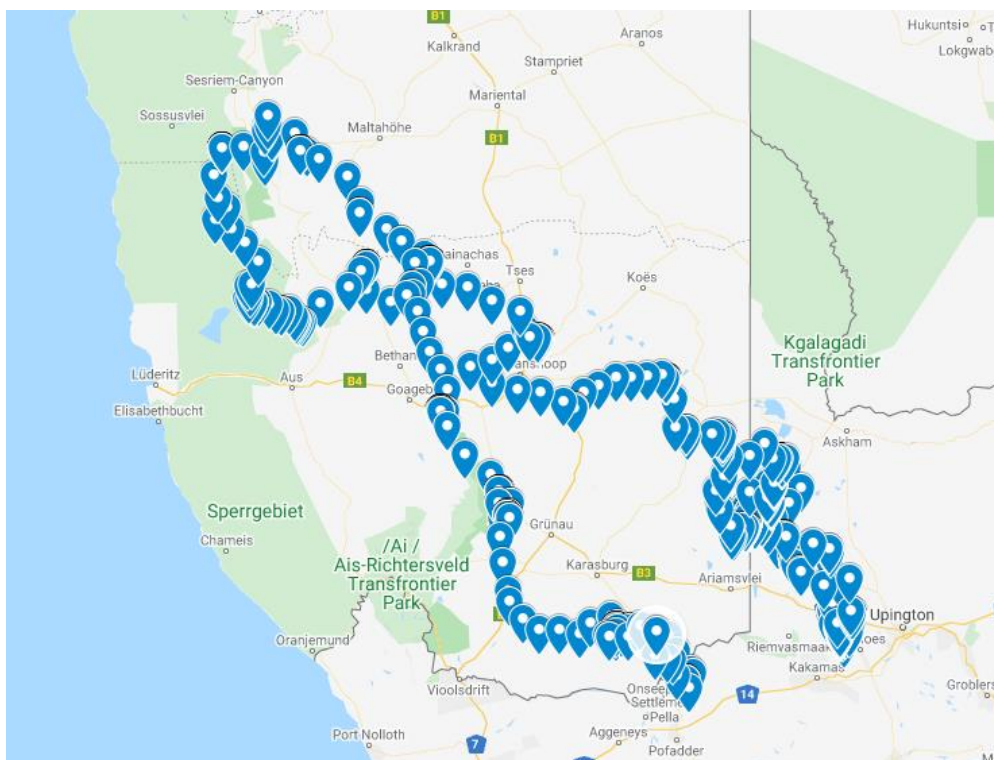
Holger Kolberg
holgerk@afol.com.na

During November and December 2018 and 2019 several Lappet-faced Vulture chicks in the Namib-Naukluft Park were fitted with tracking devices. The red star on the map is the place where the tracker was fitted. Some of the trackers take a position reading every half an hour,

others only once an hour and then there are ones that take a reading only every six hours. The distances given here are straight line measurements between the different points. The period covered in this article is from 1 November 2020 to 28 February 2021.

F272 “Amadeus” – sex ? (still to be determined), Hotsas, tracker fitted 30 December 2019. This tracker is sponsored by Namibia Tours & Safaris. Regrettably this tracker stopped functioning at the end of November 2020.

Total distance travelled since tracker was fitted: 15 899.4 km
Total distance travelled since 1 November 2020: 2 930.6 km
Largest daily movement: 234.9 km
Highest speed attained: 85.5 km/h
Highest altitude attained: 2 286 masl



F285 “Pumba” – sex ? (still to be determined), Ganab, tracker fitted 30 December 2019. This tracker is sponsored by Matiti Safaris.

Total distance travelled since tracker was fitted: 24 008.2 km

Total distance travelled since 1 November 2020: 8 945.8 km

Largest daily movement: 269.2 km

Highest speed attained: 84.5 km/h

Highest altitude attained: 3 398 masl



L727 “Oscar” – female, Kriess River, tracker fitted 9 November 2018. This tracker is sponsored by Namibia Tours & Safaris.

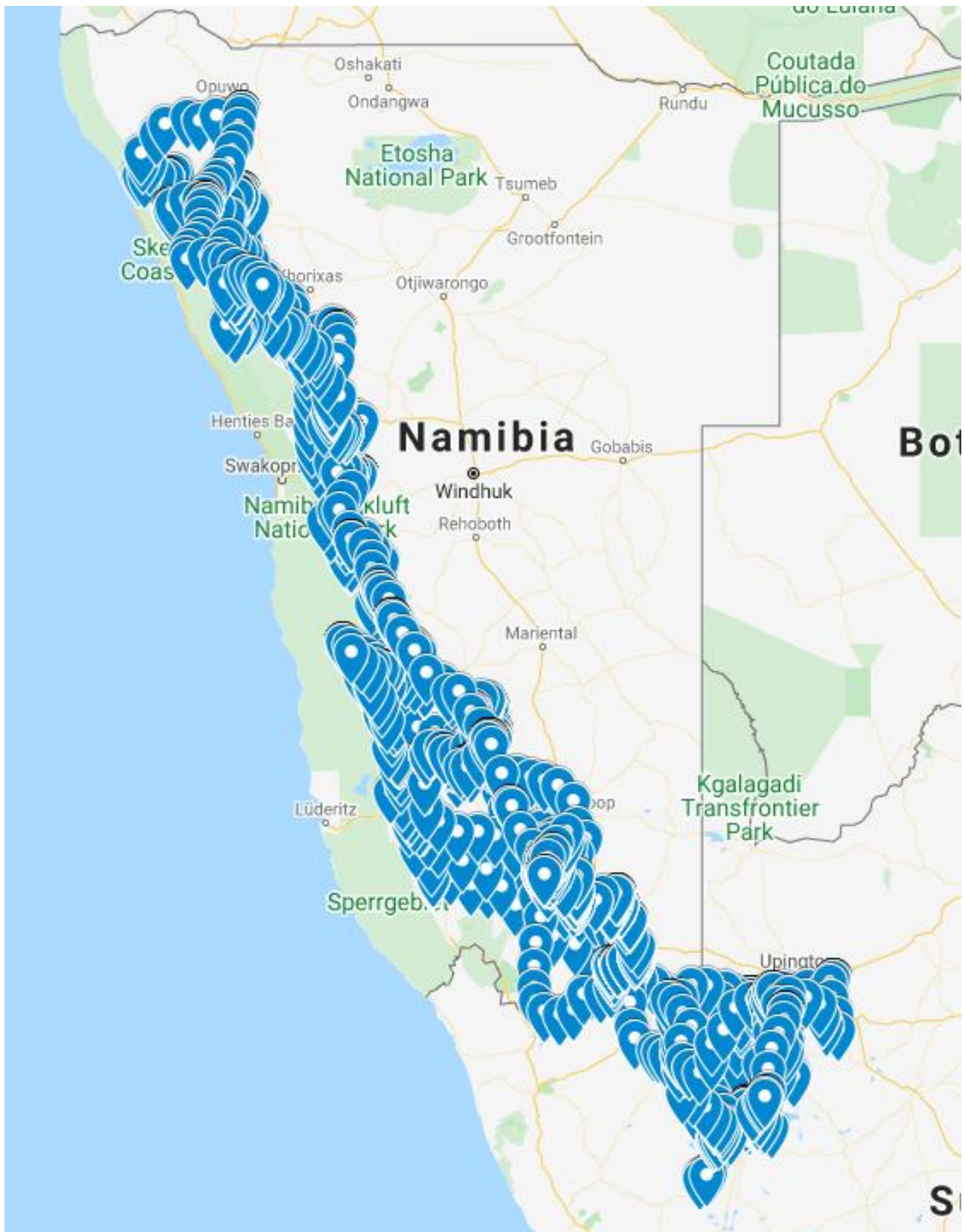
Total distance travelled since tracker was fitted: 72 178.2 km

Total distance travelled since 1 November 2020: 13 961.8 km

Largest daily movement: 319.6 km

Highest speed attained: 107.4 km/h

Highest altitude attained: 2 895 masl



L763 “Ricoffy” – sex ? (still to be determined), Elim House, tracker fitted 10 December 2019. This tracker is sponsored by Two Beards and a Saint gourmet coffee bar and bistro.

Total distance travelled since tracker was fitted: 31 606.5 km

Total distance travelled since 1 November 2020: 14 323 km

Largest daily movement: 325.3 km

Highest speed attained: 98.4 km/h
Highest altitude attained: 3 459 masl



L795 “John Travulture” – sex ? (still to be determined), Gemsbokwater, tracker fitted 8 December 2019. This tracker is sponsored by Reptile Mineral Resources and Exploration.

Total distance travelled since tracker was fitted: 32 985.3 km
Total distance travelled since 1 November 2020: 12 639.3 km
Largest daily movement: 372.1 km
Highest speed attained: 102.4 km/h
Highest altitude attained: 5 699 masl

Table 2: Species-specific names found for the birds of Namibia (See article on page 23 of this publication)

Scientific name	English	Kwangali	Rumanyo	Thimbu-kushu	Oshikwan-yama	Oshidonga	Silozi	Otjiherero	Khoekhoe-gowab	Juhoansi
Struthionidae										
Struthio camelus	Common Ostrich	mpo	mpo	mwe	omho	ompo	mpye	ombo	amis	dsuu
Numididae										
Numida meleagris	Helmeted Guineafowl		nkanga	ñanga	onghanga	onkanga	likaka	onganga		lxani / ari
Guttera pucherani	Crested Guineafowl						limpololo			
Phasianidae										
Peliperdix coqui	Coqui Francolin	sitentu								te'e
Dendroperdix sephaena	Crested Francolin	sitjindakarare	shikindakarare					ongwari ozondomba		lxai
Pternistis adspersus	Red-billed Spurfowl	siswagaragwali	likorowali / shiwali	dikorowadi				ongwari		!obo
Pternistis swainsonii	Swainson's Spurfowl							ongwari yovihua vyo mokuti		
Coturnix coturnix	Common Quail					oshimote				
Coturnix delegorguei	Harlequin Quail		lirurumbe							
Anatidae										
Dendrocygna viduata	White-faced Whistling Duck	ehilili	lihilili / lihiriri / shiyoyo	dihiriri			maiwi / meiwiye	ombaka yomurungu omuvapa		
Dendrocygna bicolor	Fulvous Whistling Duck						njeke			
Thalassornis leuconotus	White-backed Duck						kambotwe			
Plectropterus gambensis	Spur-winged Goose	esokwe	litjokwe	ditjokwi		ondjokwe	chikwe / lyokwe	ombaka yovivava vyovinyuru		gaahn
Sarkidiornis melanotos	Knob-billed Duck	nkuva	nkuva	dinguva / nguva		onkuwa / onkuya	imuyulu	ombaka yotjinyo otjiputuputu		
Alopochen aegyptiaca	Egyptian Goose		linduwe				siko	ombaka yaEgipte		

Tadorna cana	South African Shelduck							ombaka yovikongo / ombaka yozandundu		
Nettapus auritus	African Pygmy Goose						kanjeke / maiwiye			
Spatula hottentota	Blue-billed Teal							ombaka yehi rowakwena		
Spatula smithii	Cape Shoveler		lihiri					ombaka royunyo nomarama omangara		
Anas sparsa	African Black Duck							ombaka ondurozu / ombaka zorundu		
Anas undulata	Yellow-billed Duck						samba	ombaka yotjinyatjingara		
Anas platyrhynchos	Mallard				onhololo					
Anas capensis	Cape Teal							ombaka yaKapa		
Anas erythrorhyncha	Red-billed Teal					onkololo	sifuli	ombaka yoheya / ombaka yotjiserano		
Netta erythrophthalma	Southern Pochard							ombaka yokorupati rweyuva		
Oxyura maccoa	Maccoa Duck							ombaka yoviva ovisupi novizorundu		
Caprimulgidae										
Caprimulgus europaeus	European Nightjar									glanih
Caprimulgus pectoralis	Fiery-necked Nightjar			rumbamba		lumbamba				
Apodidae										
Apus affinis	Little Swift							okandiri kambepo		n!uu#hae#hae
Apus caffer	White-rumped Swift							ondiri yambepo yekonde vapa		
Musophagidae										

Corythaixoides concolor	Grey Go-away-bird	nkwe	muñwe / nkwe	mungwe / muñwe	nanghwe	nankwe / siluwewe	mukuwela / simupee / simuwe	ongurukwena	noeab	
Otididae										
Ardeotis kori	Kori Bustard		lipweyampundu	ndhinye	nakaungwa	ekakalwa / ethona		etuvangoma / etwangema		!uih / !ae!kuma
Eupodotis rueppelii	Rüppell's Korhaan								!noaes	
Lophotis ruficrista	Red-crested Korhaan		lipampa			epampa		etuva		lah / txoro
Afrotis afraoides	Northern Black Korhaan							etatewe / etewe	!gatsilgararas / gāras / xartsilgālgās	na'm
Cuculidae										
Centropus superciliosus	White-browed Coucal						mbulwenyi / mukuku / simikata			
Clamator levaillantii	Levaillant's Cuckoo							onḡera yomburombe yotjungora		
Clamator jacobinus	Jacobin Cuckoo							onḡera yomburombe		
Pteroclididae										
Pterocles gutturalis	Yellow-throated Sand-grouse						sanungana			
Pterocles burchelli	Burchell's Sand-grouse									!ao!uih
Columbidae										
Columba livia	Rock Dove			dimbutukuwa						
Columba guinea	Speckled Pigeon	haifoko		dikumbwaktji / ditjuvarupa				eṭiṭipongo / onguti pongo / onguti yutuwa	!garo+nabis	
Streptopelia semitorquata	Red-eyed Dove						simukuyu			
Streptopelia capicola	Ring-necked Dove		maranda	ñumbwakutji		elyamono / okahanda / ontendeguti		onguti ewara / onguti yaKapa yozonduso	!nū!ao+nais	
Spilopelia senegalensis	Laughing Dove	hamanku / katere	katere					ohanda	!awa!ao+nais / flōb / namakwa+nawis	

Turtur chalcospilos	Emerald-spotted Wood Dove	sitenderenkutji	shitenderekutji					onguti yotjivavize		
Oena capensis	Namaqua Dove	kambowo	kambovo				kampolwe / natangundu	onguti osurando	lhopob	lhobo / gloai
Treron calvus	African Green Pigeon	mpungu huriti / putukuwa	liputukuwa / nginga / nkutipembe	nginga			liembe / sikuluyi	onguti otjihape		
Rallidae										
Rallus caerulescens	African Rail	sipika					mumbuma			
Amaurornis flavirostra	Black Crane		tjukutjuku				mulunga mutumbala			
Gallinula chloropus	Common Moorhen					onkwinkwiti	liondongoma	ohunguriva yomewa		
Fulica cristata	Red-knobbed Coot							onhungiri vandjeo		
Gruidae										
Balearica regulorum	Grey Crowned Crane	engwangali	liñwañwali		nakandunga		liowanyi			
Grus carunculata	Wattled Crane			muhorori		eyamakalwa	libanda / mucatomo / mupuzwa			
Podicipedidae										
Tachybaptus ruficollis	Little Grebe					neumbu / neyumbo / okaneumbu / ondjumbu	kacolañasi			
Phoenicopteridae										
Phoenicopterus roseus	Greater Flamingo							kakueya		
Phoeniconaias minor	Lesser Flamingo							kakueya okañi		
Turnicidae										
Turnix sylvaticus	Common Buttonquail						kaututu	ondwezu yehezu		labu
Burhinidae										

Burhinus vermiculatus	Water Thick-knee						lundio	otjiuru otjindike tjomewa		
Burhinus capensis	Spotted Thick-knee	esivayita				ndemuteya		ipingizupauda	!gabas!nâlgob	na'm!'hauhau
Haematopodidae										
Haematopus moquini	African Oystercatcher								dōbas	
Recurvirostridae										
Himantopus himantopus	Black-winged Stilt						nalulenge			
Recurvirostra avosetta	Pied Avocet									hui'usi
Charadriidae										
Vanellus armatus	Blacksmith Lapwing		runkerenkere				ilukelekele	orukungwini oruzoronou	lganlganneb	l'ai'l'ai
Vanellus coronatus	Crowned Lapwing					luntsentsona		ongurukungwini / orukungwini rotjikorona	lgāxaris / xeixeis / xoexoes	n!aq'i
Vanellus senegallus	African Wattled Lapwing						liandanda / liandandwa			
Rostratulidae										
Rostratula benghalensis	Greater Painted-snipe									
Jacaniidae										
Microparra capensis	Lesser Jacana	nkongoro gwakambatu					naniliatandombe			
Actophilornis africanus	African Jacana		kavarambidira / nkongoro	kavarambidhira			nalumbe	onimwe mire / oyakana		
Scolopacidae										
Gallinago nigripennis	African Snipe	nkoko					longongo / luonoño			
Tringa glareola	Wood Sandpiper									g!uxamaxama
Glareolidae										

Rhinoptilus chalcopterus	Bronze- winged Courser						litololwa			
Glareola nuchalis	Rock Pratincole						muliatandombe			
Laridae										
Rynchops flavirostris	African Skimmer						mbundamushek e			
Chroicocephalu s cirrocephalus	Grey- headed Gull						sambamba			
Chroicocephalu s hartlaubii	Hartlaub's Gull								#kharigaos	
Larus dominicanus	Kelp Gull								gao!oaxas	
Hydroprogne caspia	Caspian Tern								!au!khōb	
Spheniscidae										
Spheniscus demersus	African Penguin								xoros	
Ciconiidae										
Mycteria ibis	Yellow- billed Stork	nepando								
Anastomus lamelligerus	African Openbill		litongorokofu			endongondong o		endongo rotjinya kamwaha		
Ciconia nigra	Black Stork		lindongondon go	ditongora				endongo ezorundu		
Ciconia abdimii	Abdim's Stork						mbondwanyunyi			
Ciconia ciconia	White Stork	nkumbinku mbi	nkumbinkum bi	dishingango mbe / mwombo				endongo evapa / ondera yazombahu ombapene		
Ephippiorhync hus senegalensis	Saddle- billed Stork	kandjendje	kandjendje	ghutongora			mulombwe	endongo otjikaviro		
Leptoptilos crumenifer	Marabou Stork	nyumbu	nyumbu	diguve / nyumbu				otjimbirinyam a	auraib	n!aba / n!huug!koa
Sulidae										
Morus capensis	Cape Gannet								!handas / !huni!aos / xamanis	

Phalacrocoracidae										
Microcarbo africanus	Reed Cormorant		nkorokoro							
Phalacrocorax lucidus	White-breasted Cormorant								!urilkhaib	
Phalacrocorax capensis	Cape Cormorant								ṁū-anis	
Anhingidae										
Anhinga rufa	African Darter	endeda	muṁvanda				ṁukukbiambai / ṁulindeti			
Threskiornithidae										
Threskiornis aethiopicus	African Sacred Ibis			dyaghagha		onkimbwandhila / onkumbwandhila	nasikambo			
Bostrychia hagedash	Hadada Ibis	ngoromuduv a					ṁwawawa			
Plegadis falcinellus	Glossy Ibis						ngui / ngwiwi			
Platalea alba	African Spoonbill						ṁbambakatuwa			
Ardeidae										
Nycticorax nycticorax	Black-crowned Night Heron	hakaruu gomasiku					siwa	etuva routuku		
Butorides striata	Striated Heron							etuva rotjinyo otjingirine		
Ardeola rufiventris	Rufous-bellied Heron						nuba-ṁuba			
Bubulcus ibis	Western Cattle Egret		lishingangombe	kadithangombe / kavutangombe				etuva rovinamuinyo / ondera yozongupa	gūre-aob	
Ardea cinerea	Grey Heron							etuva evahe	gurikhoeseb / gei-!noab	
Ardea melanocephala	Black-headed Heron		yenga			endoko / kandoko		etuva rotjiuru otjizorunda		
Ardea goliath	Goliath Heron						ṁbuyanda	etuva romasa		
Ardea purpurea	Purple Heron	samunkoma gomugeha					nalukapwa			

Ardea alba	Great Egret	samunkoma gomuzera					likolola / mukolola	etuva enene		
Egretta ardesiaca	Black Heron	samunkoma gomusavaga ni						etuva ezorundu		
Egretta garzetta	Little Egret							etuva okaḽiḽi		
Scopidae										
Scopus umbretta	Hamerkop	mfune	kamutuva / rufune	rufune		namutapaloya	sibengelele	otjiuru mbike		tša'antuutuu / tša'q'an
Pelecanidae										
Pelecanus onocrotalus	Great White Pelican		nekava			onyundu	liya		!ēdeharab	
Sagittariidae										
Sagittarius serpentarius	Secretary- bird	mukongo	mukongo	mukongo	hamusang a	nuukuti	mukongo	kariombo/ ombumbamen o	!āxarab / turu!khobab / tūrebe / !nora-lgaras	g'aakhoe
Accipitridae										
Elanus caeruleus	Black- winged Kite							orukoze rovivava ovizorundu		
Gypohierax angolensis	Palm-nut Vulture							onguvi ombapa		
Neophron percnopterus	Egyptian Vulture								!uri-goras	
Gyps africanus	White- backed Vulture						lipaswa			
Gyps coprotheres	Cape Vulture							onguvi yaKapa		
Trigonoceps occipitalis	White- headed Vulture							onguviyetjiuru otjivapa		
Torgos tracheliotos	Lappet- faced Vulture					elyanyama		onguvi ondorozu		
Circaetus pectoralis	Black- chested Snake Eagle							orukoze yozonoka yorukoro oruzorundu		
Circaetus cinereus	Brown Snake Eagle							orukoze yonoka / ovikurikoze yonoka		

Terathopius ecaudatus	Bateleur	sipupa	shipupa	thipungu			sibungu			nlaqu'loan
Polemaetus bellicosus	Martial Eagle	ngongo gepampa						orukoze havita / ovikurikoze havita		
Lophaetus occipitalis	Long-crested Eagle						simbukoki / siyangwa / suyangwa			
Hieraaetus wahlbergi	Wahlberg's Eagle							orukoze choni		lam
Hieraaetus pennatus	Booted Eagle							orukoze rwaKapa		
Aquila rapax	Tawny Eagle		lishambakodi / ngonga					orukoze ohonimdumbu		laoha
Aquila nipalensis	Steppe Eagle							orukoze omaheke		
Aquila verreauxii	Verreaux's Eagle	ngongo zonsavagani						orukoze ondorozu		
Aquila spilogaster	African Hawk-Eagle		likangakodi							
Kaupifalco monogrammicus	Lizard Buzzard							orukoze roviturukuhu		
Micronisus gabar	Gabar Goshawk							oharukoze okaapa / orukoze romopembany oka		
Melierax metabates	Dark Chanting Goshawk									
Melierax canorus	Pale Chanting Goshawk							orukoze orunene eruvapa		nlaeglxarice
Accipiter minullus	Little Sparrowhawk	kakodi gomununu								
Accipiter ovampensis	Ovambo Sparrowhawk							orukoze yovambo		
Accipiter melanoleucus	Black Sparrowhawk	kakodi gomusovangani								
Circus ranivorus	African Marsh Harrier						sikakuwa	orukoze romarindi waAfrika		g'ha
Circus macrourus	Pallid Harrier							orukoze oruzorundu		

Milvus migrans	Black Kite							orukuze oruzorundu		
Milvus aegyptius	Yellow-billed Kite					embiimbili / kayimbi		ombirinyama		
Haliaeetus vocifer	African Fish Eagle	mpungu	mpungu	ñwanyi		onkwalukugo		onguvi yomahundju / orukoze yomahundju	huri-!ari- khas	
Buteo augur	Augur Buzzard								!homn-arin	
Buteo rufofuscus	Jackal Buzzard							orukoze rozombandje		
Tytonidae										
Tyto alba	Western Barn Owl			dihihhi				onjimbi / otjisiwitjomah ozu	hũ#guib / sereb	nlaq'o
Strigidae										
Otus senegalensis	African Scops Owl		tukutuwi				sibengelele	okasivi kautui outiŋi		!uisa
Bubo lacteus	Verreaux's Eagle-Owl		lififita							!urukuba
Glaucidium perlatus	Pearl-spotted Owlet									!uiboog!u
Upupidae										
Upupa africana	African Hoopoe	kangungu / nduranganga		dikeke		dhepupu			!nũres	#ang#oalabutchil hae / !huma- n!ai-g!ka
Phoeniculidae										
Rhinopomastus cyanomelas	Common Scimitarbill									!x'ugaragara
Bucorvidae										
Bucorvus leadbeateri	Southern Ground Hornbill	engomba	lingomba			epumputu	lingomba	epumutu		
Bucerotidae										
Tockus rufirostris	Southern Red-billed Hornbill							etoko rotjinjotjiseran du		
Tockus leucomelas	Southern Yellow-billed Hornbill							etoko rotjinjotjingara		!oehtcu / !'horoko / tcu

Lophoceros alboterminatus	Crowned Hornbill							etoko rotjinkorone		
Lophoceros nasutus	African Grey Hornbill	munkono	munkono / nkono					etoko evahe		#hamsa
Bycanistes bucinator	Trumpeter Hornbill									!xokhoe
Coraciidae										
Coracias caudatus	Lilac- breasted Roller		shikambu				sitakela	onḡera wovanatje	kōkōias / ō- ōlnaēs	
Coracias garrulus	European Roller									tjam
Alcedinidae										
Halcyon senegalensis	Woodland Kingfisher		ntungurure					ongambura mahundju yaAngola		
Ispidina picta	African Pygmy Kingfisher						mutangalati / nengelele / sakacompole / sitaka			
Corythornis cristatus	Malachite Kingfisher						nalikwinji			
Megaceryle maxima	Giant Kingfisher	esompwanin gi	lishompwanin gi				koña			
Ceryle rudis	Pied Kingfisher		shamuningi					ongambura mahundju ombonde		
Meropidae										
Merops hirundineus	Swallow- tailed Bee- eater							onḡera yotjiongo	nīb	
Merops pusillus	Little Bee- eater						muole-ole			
Merops apiaster	European Bee-eater							onḡera yazonjutji		
Merops nubicoides	Southern Carmin Bee-eater	muhembo		rutembanday i						
Lybiidae										
Tricholaema leucomelas	Acacia Pied Barbet							ombonde / ombendomuti	kurib toas	
Lybius torquatus	Black- collared Barbet						sifula-nyundo			

Indicatoridae										
Indicator minor	Lesser Honeyguide									loaq
Indicator indicator	Greater Honeyguide			ndegha						
Picidae										
Campethera abingoni	Golden-tailed Woodpecker			mbangura				ombetamiti		
Chloropicus namaquus	Bearded Woodpecker		djaradjara							
Dendropicos fuscescens	Cardinal Woodpecker							ongongoramuti	hailgō!gōseb	
Falconidae										
Polihierax semitorquatus	Pygmy Falcon							orukoze okaṭṭi		
Falco naumanni	Lesser Kestrel							orukoze oruserandu / orukoze oruserandu oruṭṭi		
Falco rupicolus	Rock Kestrel							orukoze romawe		
Falco rupicoloides	Greater Kestrel							orukoze orunene		
Falco dickinsoni	Dickinson's Kestrel							orukoze oruzandu rovyoze		
Falco chicquera	Red-necked Falcon							orukoze rosengu serandu		#hangma
Falco amurensis	Amur Falcon							orukoze rozombaze ozeserandu rokomuhuka		
Falco peregrinus	Peregrine Falcon									!ao
Psittacidae										
Poicephalus meyeri	Meyer's Parrot		poro				sicikwele			tsaqni

Psittaculidae										
Agapornis roseicollis	Rosy-faced Lovebird					ehwilihwili		onḡera tḡapolise		
Malaconotidae										
Tchagra senegalus	Black-crowned Tchagra						sasibo			
Laniarius major	Tropical Boubou									kxaruḡi!aian
Laniarius atrococcineus	Crimson-breasted Shrike	etwakura	litwakura	ditwa				orowa / otḡihuna maverere	naorab	
Laniidae										
Urolestes melanoleucus	Magpie Shrike	muruli	muruli							
Eurocephalus anguithimens	Southern White-crowned Shrike		kekeya							
Lanius collurio	Red-backed Shrike		nankugho							
Oriolidae										
Oriolus larvatus	Black-headed Oriole			ndurunyunyi						
Dicruridae										
Dicrurus adsimilis	Fork-tailed Drongo	ntene	ntene			ontene	mutengumusha mba		lḡauseb / ḡūdarub / piridarub	n!hoce
Monarchidae										
Terpsiphone viridis	African Paradise Flycatcher		karuvadi	nderendere			silusila-sila			
Corvidae										
Corvus capensis	Cape Crow		likorova			edhilakola		otḡikwarikoko / otḡikwarikwak wa	†nūḡorab / †nūnāeb / †nūnāegorab	
Corvus albus	Pied Crow		likombanturo				lihakabe	otḡikuara	!uril!khaigora b	
Alaudidae										

Eremopterix leucotis	Chestnut-backed Sparrow-Lark					nambololola				
Pycnonotidae										
Pycnonotus nigricans	African Red-eyed Bulbul							okadubombi / okaotamea		#umakoqrikoqri
Pycnonotus tricolor	Dark-capped Bulbul						cimpolyo / simplyo		#aromüeb / #aromükuieb / kamtrokuieb	
Hirundinidae										
Riparia paludicola	Brown-throated Martin						katwele			
Ptyonoprogne fuligula	Rock Martin							ondiri romawe		
Cecropis cucullata	Greater Striped Swallow							ondiri yotjari totungora otunene		
Macrosphenidae										
Sylvietta rufescens	Long-billed Crombec					ndulumakunde				
Cisticolidae										
Prinia flavicans	Black-chested Prinia							okahonatjiunda		
Leiiothrichidae										
Turdoides jardineii	Arrow-marked Babbler									g'o'oro
Turdoides bicolor	Southern Pied Babbler							onqera otjiapa		g!kauce
Sturnidae										
Lamprotornis nitens	Cape Starling					oshidhilu			#khainab	
Lamprotornis chalybaeus	Greater Blue-eared Starling		ndiru				ndilu			

Lamprotornis australis	Burchell's Starling		ndirure						Inuwub / arurub	
Buphagidae										
Buphagus africanus	Yellow-billed Oxpecker		kamugcara				cabañombe			
Buphagus erythrorhynchus	Red-billed Oxpecker			ghuthe						
Muscicapidae										
Cercotrichas coryphoeus	Karoo Scrub Robin									g!ani!ae!aece
Cercotrichas paena	Kalahari Scrub Robin								gapitsirob	
Cercotrichas leucophrys	White-browed Scrub Robin	eherekete	liherekete	munembo		shikotoimba				
Saxicola torquatus	African Stonechat						sitengu			!oro
Oenanthe pileata	Capped Wheatear	kazinganzwi							iab	
Nectariniidae										
Chalcomitra senegalensis	Scarlet-chested Sunbird						masinja	otjiondaosond u		
Cinnyris talatala	White-bellied Sunbird									goh-tzama
Passeridae										
Passer motitensis	Great Sparrow							oyatuhere		!oqmihia / joaqi
Passer melanurus	Cape Sparrow					evalelanganda		oyatuhere rwaKapa		
Ploceidae										
Bubalornis niger	Red-billed Buffalo Weaver								!khuis / au-anis	karikari / n!angkarikari
Plocepasser mahali	White-browed Sparrow-Weaver							onɔera yovikokotua		!aun-!auhn

Philetairus socius	Sociable Weaver							onḡera yomuhoko	!hanageti / !hanati	
Sporopipes squamifrons	Scaly-feathered Weaver					naangongwa		okaryande		lari / g'uilari
Ploceus intermedius	Lesser Masked Weaver							okarumboira okaṡiṡi		
Ploceus velatus	Southern Masked Weaver							okarumboira	!khanikexaus / !gis / ṡā-anis / ṡās / ṡorobes	
Quelea quelea	Red-billed Quelea		mandjunge	dimba dyo thiyunyi		eyandja				
Euplectes afer	Yellow-crowned Bishop						lishewa			
Euplectes orix	Southern Red Bishop	gomugeha	kambara	ditwamukura						g!aun-laqnaxxai
Estrildidae										
Pytilia melba	Green-winged Pytilia								!gibes	
Amadina erythrocephala	Red-headed Finch	kafilita							ṡgāges / ṡhāus	
Lagonosticta senegala	Red-billed Firefinch						kanwele / nakanwele			
Uraeginthus angolensis	Blue Waxbill	katjikilili gomuburau	kandjerendjen dje			naashiishii				
Uraeginthus granatinus	Violet-eared Waxbill									l'uri
Estrilda astrild	Common Waxbill					naakwalukula				
Estrilda erythronotos	Black-faced Waxbill	siguye	shiguye			nalukula				
Ortygospiza atricollis	Quailfinch	mukadi gonon								tintin
Lonchura cucullata	Bronze Mannikin						kambalamatende			
Viduidae										
Vidua macroura	Pin-tailed Whydah						silucila-cila			
Vidua regia	Shaft-tailed Whydah							omuhona wosondera		

Vidua paradisaea	Long-tailed Paradise Whydah									da'atzama
Motacillidae										
Motacilla capensis	Cape Wagtail					naatooto	nalungwana			
Motacilla aguimp	African Pied Wagtail		kamukombo				kashela			
Macronyx ameliae	Rosy-throated Longclaw						nalutota			
Anthus vaalensis	Buffy Pipit									gi'o'ore
Fringillidae										
Crithagra atrogularis	Black-throated Canary	kandingo gomonondundu				namukuku				
Crithagra mozambica	Yellow-fronted Canary	kamukombo nsense								

Membership Fees 2021

African Birdlife, the magazine published by BirdLife South Africa can be ordered through the Namibia Bird Club. A year's subscription (6 issues) of African Birdlife currently costs N\$ 330.00

Category (excluding African Birdlife subscription)

Single member	N\$ 170
Family membership (husband, wife, children)	N\$ 220
Junior member (all scholars and students).....	N\$ 90
Pensioner	N\$ 120
Corporate membership.....	N\$ 700

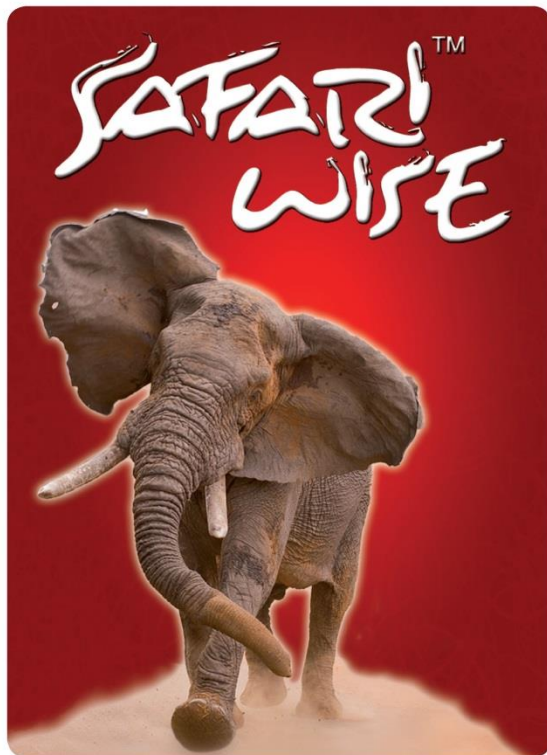
Namibia Bird Club Committee 2020/21

Gudrun Middendorff (Chairperson)	Tel: 081 240 3635
Gertrud Piek (Treasurer)	Tel: 061 24 1838 (home)
Ellen Gudde (Secretary)	Tel: 081 248 7362
Franz Klein (Website Manager)	Tel: 081 210 1221 email: franzaklein@gmail.com
Holger Kolberg (Editor Lanioturdus)	Tel: 081 129 5163 email: holgerk@afol.com.na
Sonja Bartlewski (Member)	Tel: 081 149 2313
Anita Zimny (Member)	Tel: 081 280 2527

Advertising in Lanioturdus

We will accept advertisements to be published in Lanioturdus at the following rates:

Full page	N\$ 300
Half page	N\$ 150
Quarter page.....	N\$ 75
Small advertisement.....	N\$ 35



Africa Waiting for You!

- Personalised: to your needs
- Small Groups: to keep it intimate
- Special Interests: as per your requests

www.safariwise.com.na

www.birdwatching.com.na



SAFARIWISE • PO Box 2796 • Swakopmund • Namibia • T/F: + 264 64 40 5220 • M: + 264 81 124 9391 • E: neil@safariwise.com.na • S: safariwisenamibia

SAUVAGE
AFRICAN JOURNEYS



Instructions to Authors

Lanioturdus is a journal dedicated to birds and birding. Although the journal's primary focus is on Namibia, articles from other geographical parts of the globe will also be considered for publication. Authors should use common and scientific names of birds as published by the Namibia Bird Records Committee (www.the-eis.com/namibia-bird-list.php) or, for birds not on the Namibian list, the latest published IOC list (www.worldbirdnames.org). Note that bird names are capitalised e.g. House Sparrow whereas reference to a group is not e.g. we saw a flock of sparrows. Text should be submitted as a MS Word document. Photos, maps and figures should be sent as separate jpeg images, graphs as MS Excel charts or jpeg images and tables as MS Word or Excel documents. Please indicate in the article text where these should be placed. Literature referred to in the text of an article must be arranged chronologically as follows: Adam (2000); Adam (2000a, 2000b); Adam and Eve (2003); Adam (2000, 2003); (Adam 2000, Albert 2001). For more than two authors, use the first author's name followed by *et al.* (e.g. Adam *et al.* 2007). At the end of the article provide a full list of references sorted alphabetically and then chronologically, using the following format:

Payne, R.B. and Payne, L.L. 1997. Field observations, experimental design, and the time and place of learning in bird songs. In: C. Snowdon and M. Hausberger (eds.), *Social influences on vocal development*. Cambridge University Press, Cambridge, UK. pp 57–84.

Urban, E.K., Fry, C.H. and Keith, S. (eds.). 1997. *The birds of Africa*, Vol. 5. Academic Press, London, UK.

Vergara, P., Aguirre, J.I. and Fernández-Cruz, M. 2007. Arrival date, age and breeding success in White Stork *Ciconia ciconia*. *Journal of Avian Biology* **38**: 573–579.

du Plessis, M.A. 2005. Green Woodhoopoe. In: Hockey, P.A.R., Dean, W.R.J. and Ryan, P.G. (eds.), *Roberts birds of southern Africa* (7th Edn). Trustees of the John Voelcker Bird Book Fund, Cape Town, RSA. pp 162–164.